

## AMMI Canada Medical Student Research Proposal

**Project:** *Corynebacterium diphtheriae* infections in an impoverished, inner-city patient population: epidemiology and mechanisms of antibiotic resistance

**Applicant:** Jason Zou, Max Rady College of Medicine, University of Manitoba

**Supervisors:** Dr. Christopher Lowe and Dr. Marc Romney, Department of Pathology and Laboratory Medicine, University of British Columbia

### Background

*Outbreaks and severe disease associated with non-toxigenic Corynebacterium diphtheriae are increasingly described worldwide, including Canada.*

Globally, routine vaccination with diphtheria toxoid has greatly reduced cases of diphtheria due to infection by toxigenic *Corynebacterium diphtheriae* (1). Despite this, there have been numerous outbreaks of diphtheria reported globally since 2000 (2). Furthermore, the vaccine does not protect against non-toxigenic *C. diphtheriae*, which has been increasingly recognized as an emergent pathogen, especially among urban, impoverished populations in the developed world (3). Reports of non-toxigenic *C. diphtheriae* causing significant disease in the form of cutaneous diphtheria, as well as invasive infections such as bacteremia and endocarditis, are occurring with greater frequency (4-10).

*Interpretive breakpoints for susceptibility testing have been updated, but clinical implications of isolates previously characterized as susceptible now reported intermediate are uncertain.*

Penicillin and erythromycin are considered first-line antibiotics used to treat diphtheria (11). In recent years, a limited number of *C. diphtheriae* cases from Canada, the US, and the UK have exhibited resistance to penicillin and other conventional antimicrobials (10, 12, 13). The Clinical & Laboratory Standards Institute (CLSI) recently lowered the penicillin susceptible breakpoint from  $\leq 1$  mg/L to  $\leq 0.12$  mg/L. This has resulted in a reclassification of many *C. diphtheriae* isolates from being classified as penicillin susceptible to penicillin intermediate (14). However, clinical evidence of penicillin resistant *C. diphtheriae* is still limited to isolated case reports; moreover, it is unclear whether these cases reflect overall trends in the prevalence of resistant *C. diphtheriae*. Treatment options for penicillin-intermediate *C. diphtheriae* are limited by gastrointestinal side effects, risk of *C. difficile* infection, and broad spectrum antibiotic exposure for erythromycin, clindamycin and vancomycin, respectively. Therefore, there is a need for larger-scale susceptibility data to help establish the underlying epidemiology of resistant *C. diphtheriae* strains, and the potential implications of resistance on the interpretation of *C. diphtheriae* susceptibility testing in the clinical microbiology laboratory.

*There is an opportunity to better understand the epidemiology of non-toxigenic C. diphtheriae resistance in a Canadian centre where this emerging pathogen is prevalent.*

Our institution (St. Paul's Hospital, Vancouver BC) serves an impoverished, inner-city community in which cutaneous *C. diphtheriae* infection is not uncommon (3, 4, 15). Unlike many other microbiology laboratories in Canada, this has allowed for the accumulation of a significant number of clinical *C. diphtheriae* isolates over recent years. We aim to use this repository of strains to study the prevalence and evolving patterns of antibiotic susceptibility among *C. diphtheriae* isolates over the span of several years, as well as characterize potential mechanisms of antibiotic resistance.

## Objectives

1. Evaluate the prevalence and patterns of antibiotic susceptibility among *C. diphtheriae* isolates collected over several years at an inner-city, tertiary care hospital.
2. Identify mechanisms of resistance through whole-genome sequencing (WGS).
3. Compare phenotypic interpretations of resistance (14, 16) with genotypic resistance analysis.
4. Determine clonality of *C. diphtheriae* isolates through both multilocus sequence typing (MLST) and WGS subtyping.
5. Determine toxigenic status of *C. diphtheriae* isolates by WGS and compare results with genotypic and phenotypic toxin testing performed by the national reference laboratory.

## Methods

### *C. diphtheriae* isolates

Approximately 100 *C. diphtheriae* isolates recovered from patients presenting to our hospital over the past 5 years will be included in this study. Isolates were recovered primarily from wound and blood cultures, and confirmed as *C. diphtheriae* as previously outlined (4). All isolates in this study were non-toxigenic as determined by the reference laboratory.

### *Antimicrobial susceptibility testing*

Susceptibility testing will be carried out on *C. diphtheriae* isolates utilizing both Etest and agar dilution. Testing for penicillin, clindamycin, erythromycin and vancomycin will be completed based on CLSI M-45 guidelines (14). Susceptibility of isolates will be interpreted utilizing both the 2010 (16) and 2015 (14) breakpoints.

### *Genomic analysis*

Whole-genome sequencing will be performed on all study isolates with next-generation sequencing (MiSeq, Illumina, San Diego, CA). Library preparation will be done using the KAPA Library HyperPlus Kit (Roche Sequencing, Pleasanton, CA).

Sequencing data will be analyzed for:

- MLST typing
- Identification of resistance markers for penicillin, macrolides and clindamycin
- Presence of diphtheria toxin gene

## Clinical Significance

*C. diphtheriae* continues to cause significant invasive and cutaneous disease, for which penicillin and erythromycin remain the mainstays of therapy. Despite isolated reports, the prevalence of antibiotic-resistant *C. diphtheriae* remains unknown. To our knowledge, this study will be the first to utilize a large sample of *C. diphtheriae* isolates to estimate the prevalence of antibiotic resistance, as well as monitor developing trends in resistance over time. These results may have implications for interpretation and reporting of *C. diphtheriae* susceptibility testing within clinical microbiology laboratories and, more broadly, the choice of first-line antibiotic (penicillin vs. erythromycin) for clinicians treating *C. diphtheriae* infections.

## References

1. Galazka A. The changing epidemiology of diphtheria in the vaccine era. *The Journal of infectious diseases*. 2000;181 Suppl 1:S2-9.
2. Sangal V, Hoskisson PA. Evolution, epidemiology and diversity of *Corynebacterium diphtheriae*: New perspectives on an old foe. *Infection, genetics and evolution : journal of molecular epidemiology and evolutionary genetics in infectious diseases*. 2016;43:364-70.
3. Lowe CF, Bernard KA, Romney MG. Cutaneous diphtheria in the urban poor population of Vancouver, British Columbia, Canada: a 10-year review. *J Clin Microbiol*. 2011;49(7):2664-6.
4. Romney MG, Roscoe DL, Bernard K, Lai S, Efstratiou A, Clarke AM. Emergence of an invasive clone of nontoxigenic *Corynebacterium diphtheriae* in the urban poor population of Vancouver, Canada. *J Clin Microbiol*. 2006;44(5):1625-9.
5. Hirata Jr R, Pereira GA, Filardy AA, Gomes DL, Damasco PV, Rosa AC, et al. Potential pathogenic role of aggregative-adhering *Corynebacterium diphtheriae* of different clonal groups in endocarditis. *Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas*. 2008;41(11):986-91.
6. Zasada AA, Zaleska M, Podlasin RB, Seferynska I. The first case of septicemia due to nontoxigenic *Corynebacterium diphtheriae* in Poland: case report. *Annals of clinical microbiology and antimicrobials*. 2005;4:8.
7. Zasada AA. Nontoxigenic highly pathogenic clone of *Corynebacterium diphtheriae*, Poland, 2004-2012. *Emerg Infect Dis*. 2013;19(11):1870-2.
8. Clinton LK, Bankowski MJ, Shimasaki T, Sae-Ow W, Whelen AC, O'Connor N, et al. Culture-negative prosthetic valve endocarditis with concomitant septicemia due to a nontoxigenic *Corynebacterium diphtheriae* biotype *gravis* isolate in a patient with multiple risk factors. *J Clin Microbiol*. 2013;51(11):3900-2.
9. Wojewoda CM, Koval CE, Wilson DA, Chakos MH, Harrington SM. Bloodstream infection caused by nontoxigenic *Corynebacterium diphtheriae* in an immunocompromised host in the United States. *J Clin Microbiol*. 2012;50(6):2170-2.
10. Fricchione MJ, Deyro HJ, Jensen CY, Hoffman JF, Singh K, Logan LK. Non-Toxicogenic Penicillin and Cephalosporin-Resistant *Corynebacterium diphtheriae* Endocarditis in a Child: A Case Report and Review of the Literature. *Journal of the Pediatric Infectious Diseases Society*. 2014;3(3):251-4.
11. Wilson AP. Treatment of infection caused by toxigenic and non-toxicogenic strains of *Corynebacterium diphtheriae*. *The Journal of antimicrobial chemotherapy*. 1995;35(6):717-20.
12. Mina NV, Burdz T, Wiebe D, Rai JS, Rahim T, Shing F, et al. Canada's first case of a multidrug-resistant *Corynebacterium diphtheriae* strain, isolated from a skin abscess. *J Clin Microbiol*. 2011;49(11):4003-5.
13. FitzGerald RP, Rosser AJ, Perera DN. Non-toxicogenic penicillin-resistant cutaneous *C. diphtheriae* infection: a case report and review of the literature. *Journal of infection and public health*. 2015;8(1):98-100.
14. CLSI. *Methods for Antimicrobial Dilution and Disk Susceptibility Testing of Infrequently Isolated or Fastidious Bacteria*. 3rd ed. CLSI guideline M45. Wayne, PA: Clinical and Laboratory Standards Institute; 2015.
15. Cockcroft WH, Boyko WJ, Allen DE. Cutaneous infections due to *Corynebacterium diphtheriae*. *Canadian Medical Association journal*. 1973;108(3):329-31.
16. CLSI. *Methods for Antimicrobial Dilution and Disk Susceptibility Testing of Infrequently Isolated or Fastidious Bacteria; Approved Guideline-Second Edition*. CLSI document M45-A2. Wayne, PA: Clinical and Laboratory Standards Institute; 2010.