

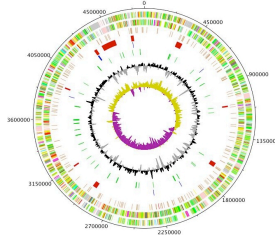
Diagnostics Arsenal for Emerging Infectious Diseases

Niaz Banaei, MD
Stanford University

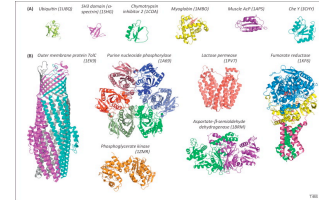
Overview

- Accurate diagnostics in 21st century
- Diagnostics for emerging pathogens
 - Ebola and Zika
 - Carbapenem-Resistant Enterobacteriaceae (CRE)
 - Malaria

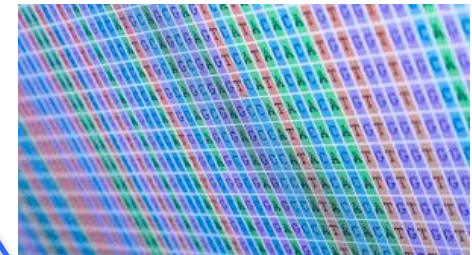
Recipe for POC Diagnostic Development



Genomics/
Proteomics Databases



Molecular
Diagnostic
Platforms



Bioinformatics
Tools

Diagnostics No Longer a Limiting Factor

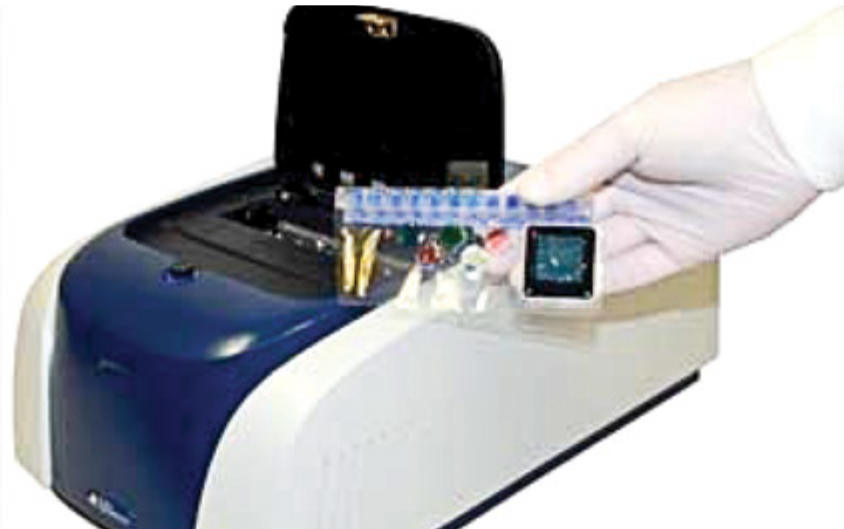
GeneXpert Omni, Cepheid



IMMY



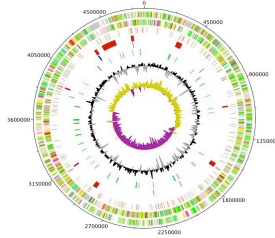
Biofire, bioMérieux



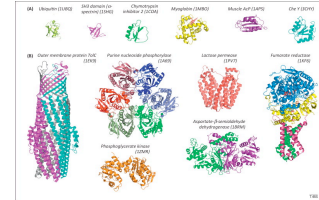
Cobas Liat System, Roche



Recipe for POC Diagnostic Development



Genomics/
Proteomics Databases



Molecular
Diagnostic
Platforms



Bioinformatics
Tools

Ebola global response was 'too slow', say health experts

🕒 23 November 2015 | Health



Healthcare workers were among those most at risk of catching Ebola

Will Ebola change the game? Ten essential reforms before the next pandemic. The report of the Harvard-LSHTM Independent Panel on the Global Response to Ebola



Suerie Moon, Devi Sridhar, Muhammad A Pate, Ashish K Jha, Chelsea Clinton, Sophie Delaunay, Valnora Edwin, Mosoka Fallah, David P Fidler, Laurie Garrett, Eric Goosby, Lawrence O Gostin, David L Heymann, Kelley Lee, Gabriel M Leung, J Stephen Morrison, Jorge Saavedra, Marcel Tanner, Jennifer A Leigh, Benjamin Hawkins, Liana R Woskie, Peter Piot

“Although scientists had identified the Ebola nearly four decades earlier and basic research had advanced understanding of the disease, Ebola was not an attractive target for industry investment in research and development, nor was it high on the public health research agenda.”

Will Ebola change the game? Ten essential reforms before the next pandemic. The report of the Harvard-LSHTM Independent Panel on the Global Response to Ebola



Suerie Moon, Devi Sridhar, Muhammad A Pate, Ashish K Jha, Chelsea Clinton, Sophie Delaunay, Valnora Edwin, Mosoka Fallah, David P Fidler, Laurie Garrett, Eric Goosby, Lawrence O Gostin, David L Heymann, Kelley Lee, Gabriel M Leung, J Stephen Morrison, Jorge Saavedra, Marcel Tanner, Jennifer A Leigh, Benjamin Hawkins, Liana R Woskie, Peter Piot

Recommendation 7: Establish a global facility to finance, accelerate, and prioritise research and development.

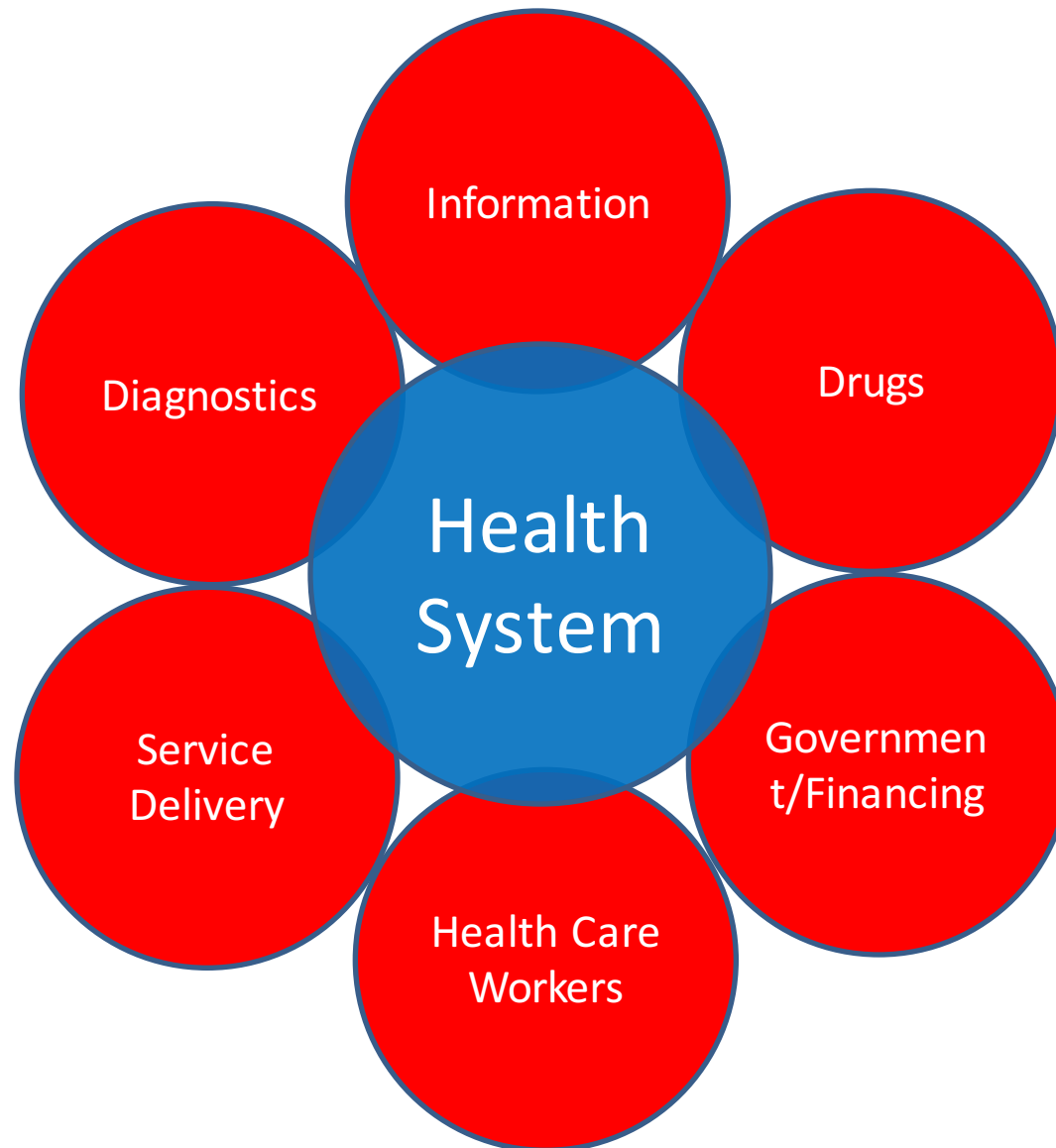
The UN Secretary General and the WHO Director-General should convene in 2016 a high-level summit of public, private, and not-for-profit research funders to establish a global financing facility for research and development for health technology relevant for major disease outbreaks. The facility would support manufacturing, research, and development for drugs, vaccines, diagnostics, and other non-pharmaceutical supplies (such as personal protective equipment) where the commercial market does not offer appropriate incentives. For known pathogens, the facility could invest in bringing candidate drugs, vaccines, technology platforms, and other relevant products through proof of concept, phase 1, and phase 2 testing in humans, so that they are ready for wider testing, manufacturing, and distribution when an outbreak strikes. During an outbreak the facility would rapidly mobilise finance for priority research and development projects, such as diagnostics for novel pathogens.

WHO Lists Cepheid's Xpert Ebola as Eligible for Procurement to Ebola Affected Countries

Xpert Ebola listed in World Health Organization Emergency Use Assessment and Listing program for diagnostic products in the context of global health emergency



Diagnostics are One Component of Health System



Laboratory Capacity is Lacking in Endemic Countries



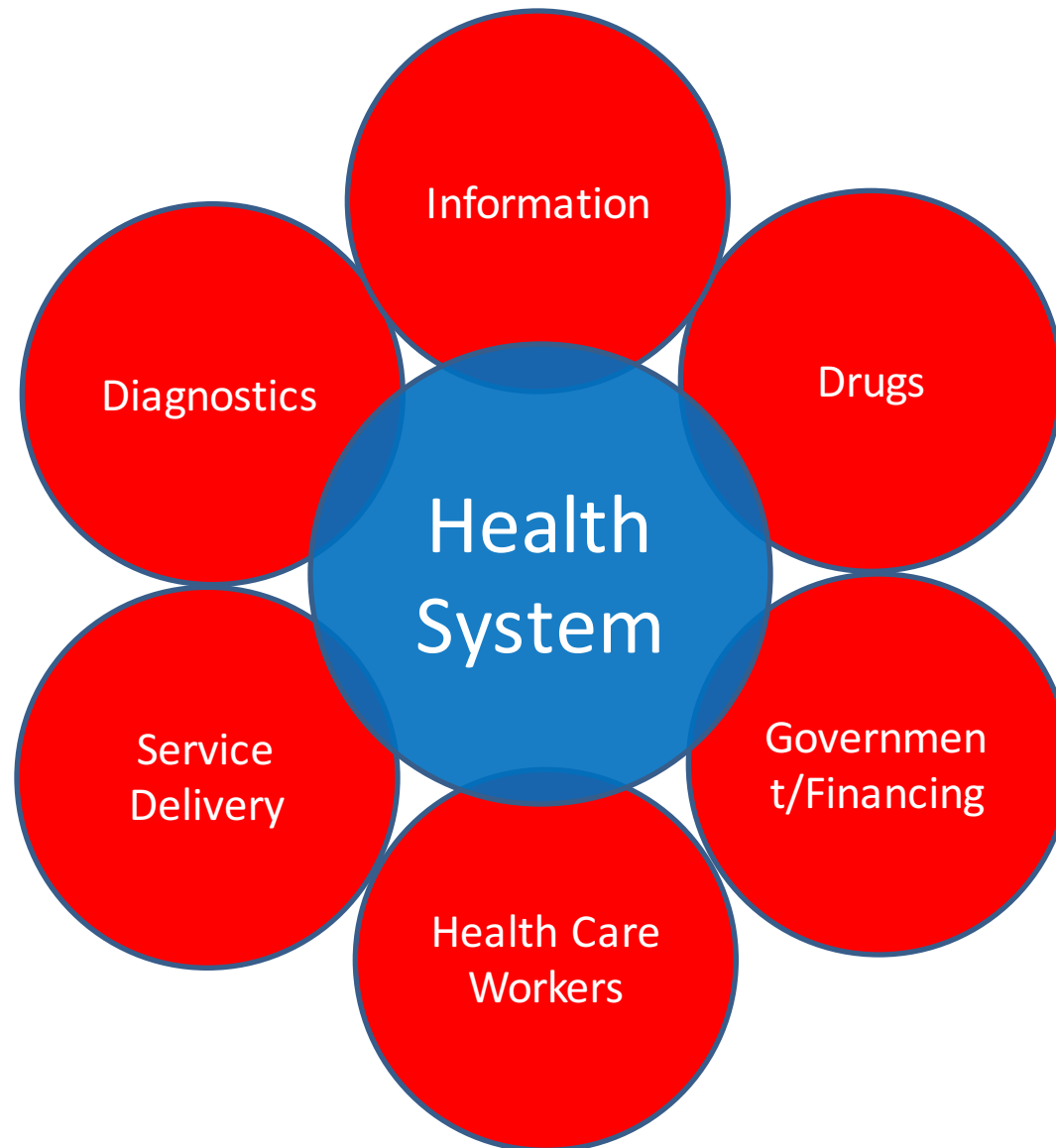
Outbreak Response Requires a Health System



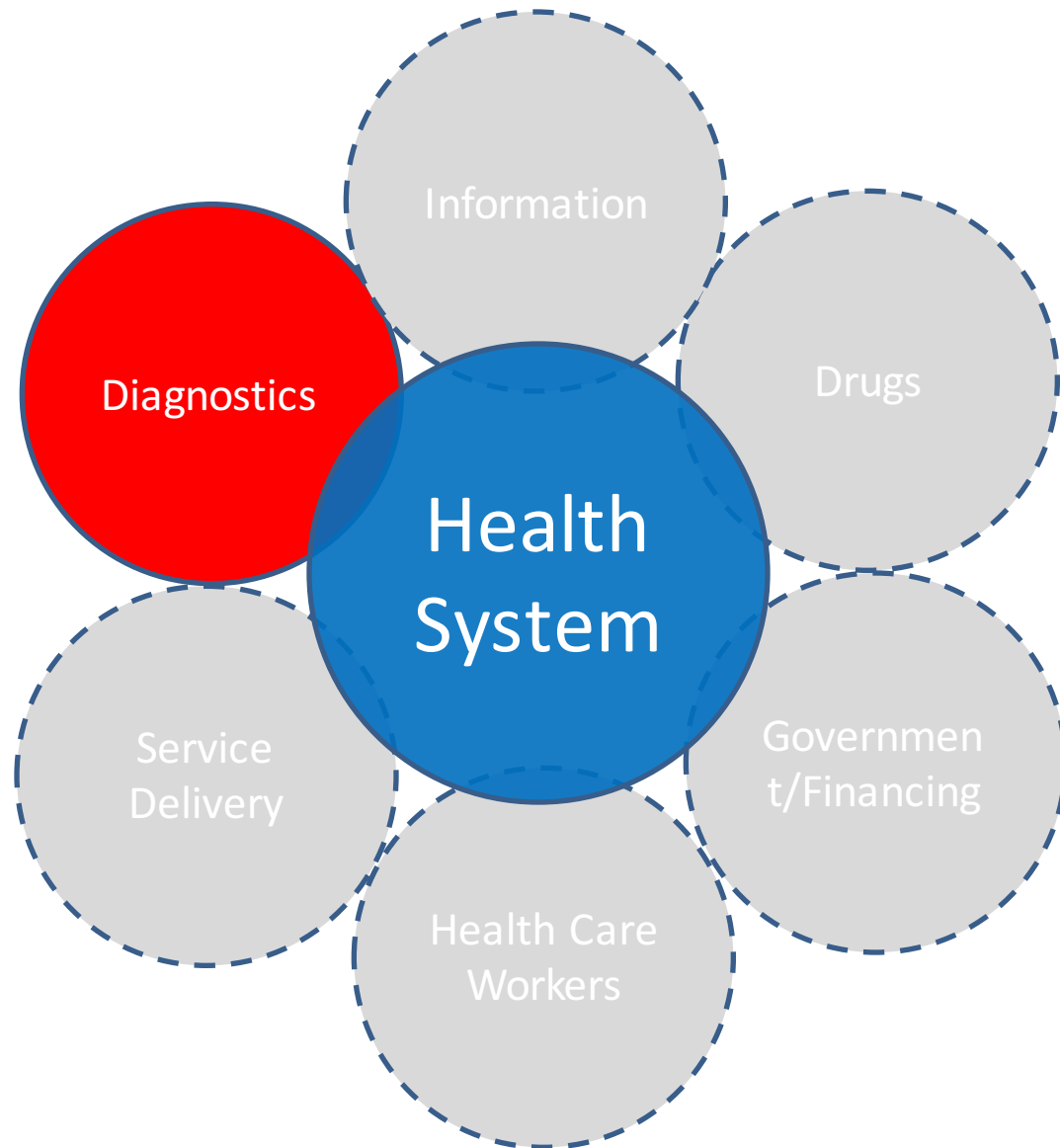
Laboratory Training is Needed in Endemic Countries



Diagnostics are One Component of Health System



Diagnostics are One Component of Health System



Public Health England laboratory in Port Loko, Sierra Leone



Credit: Jana Broadhurst

Burial team collecting dead bodies from a treatment center at Koidu Government Hospital where many healthcare workers had fallen victim due to inadequate infection control measures.



Credit: Jana Broadhurst

Delivering blood samples by motorbike to the Public Health England field diagnostic laboratory in Port Loko, Sierra Leone



Credit: Jana Broadhurst

Overview

- Accurate diagnostics in 21st century
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Point Of Care Ebola Diagnostics



Corgenix ReEBOV Antigen Rapid Test

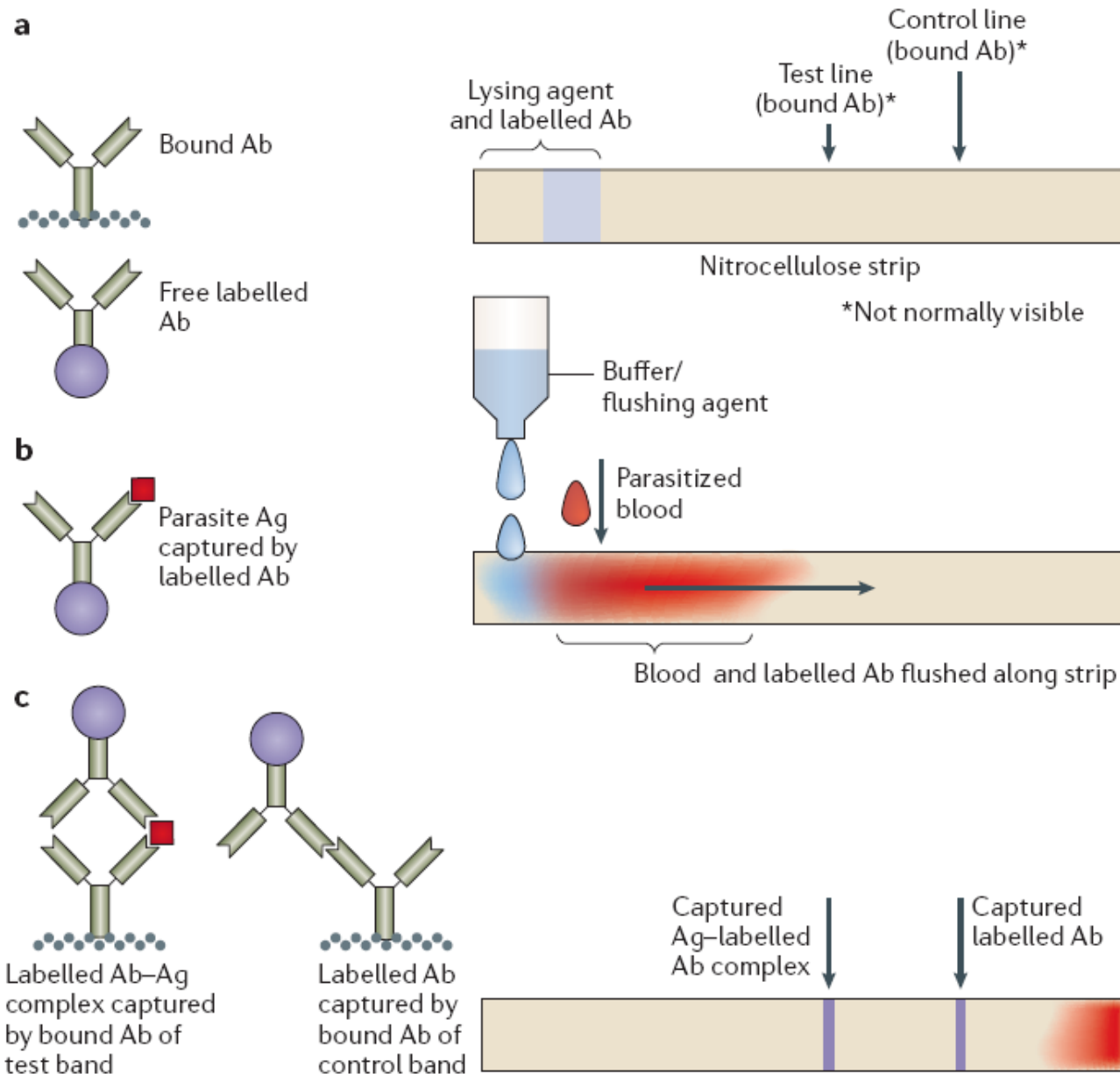


Method: Dipstick immunoassay

Target: Detecting VP40 matrix protein

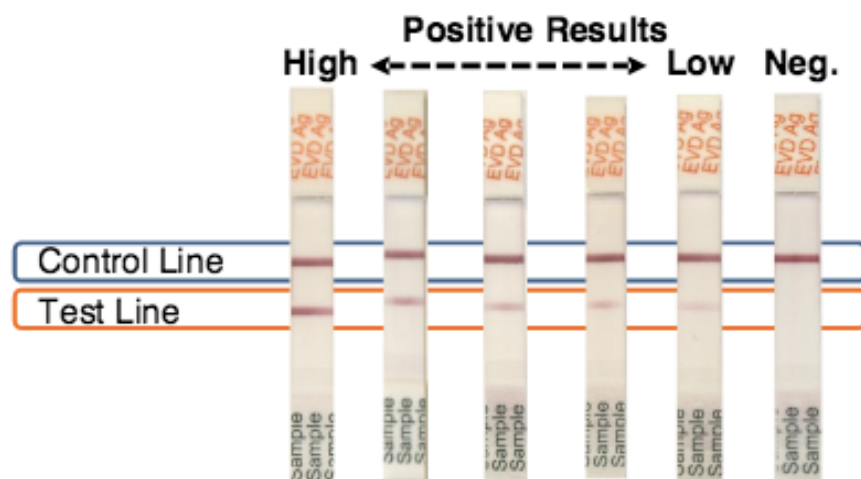
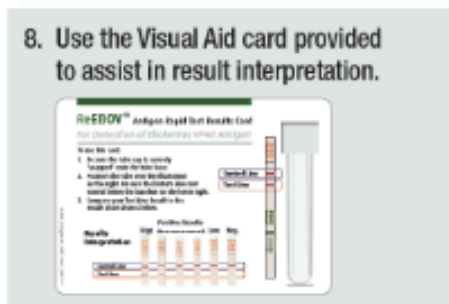
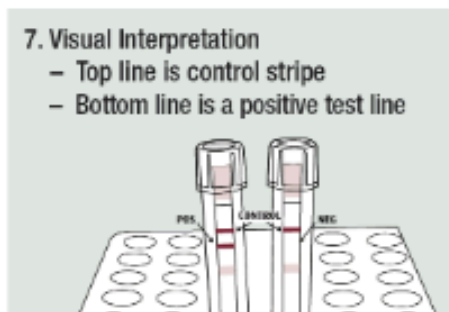
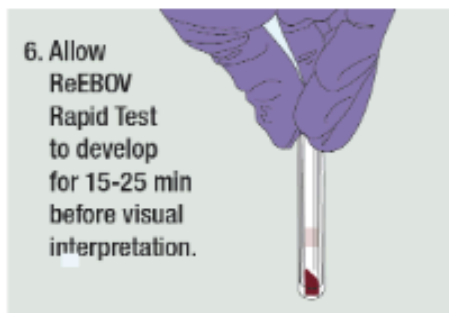
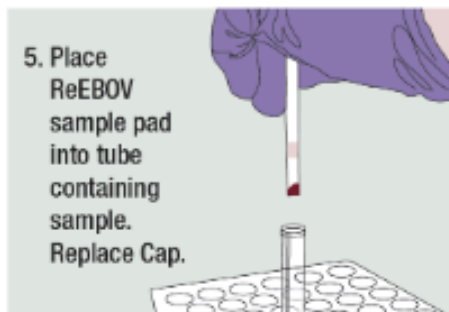
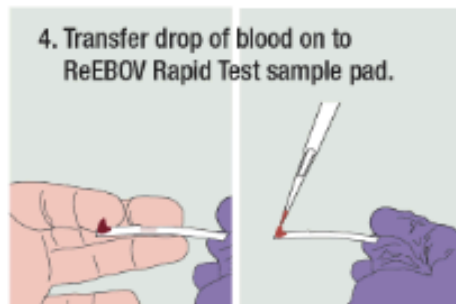
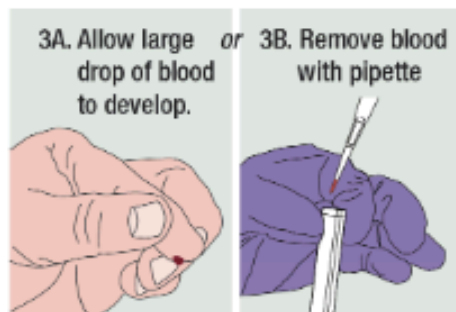
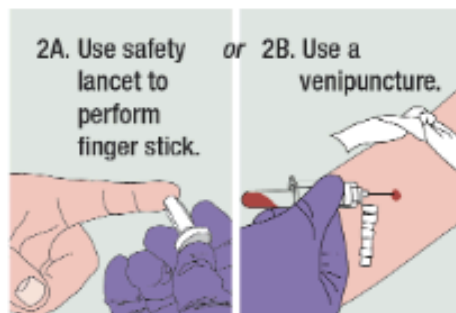
TAT: 15 to 25 minutes

Lateral flow immunochromatographic assay



ReEBOV™ Antigen Rapid Test Instructions

For Detection of Ebolavirus VP40 Antigen



ReEBOV Antigen Rapid Test kit for point-of-care and laboratory-based testing for Ebola virus disease: a field validation study

Mara Jana Broadhurst, John Daniel Kelly, Ann Miller, Amanda Semper, Daniel Bailey, Elisabetta Groppelli, Andrew Simpson, Tim Brooks, Susan Hula, Wilfred Nyoni, Alhaji B Sankoh, Santigi Kanu, Alhaji Jalloh, Quy Ton, Nicholas Sarchet, Peter George, Mark D Perkins, Betsy Wonderly, Megan Murray, Nira R Pollock

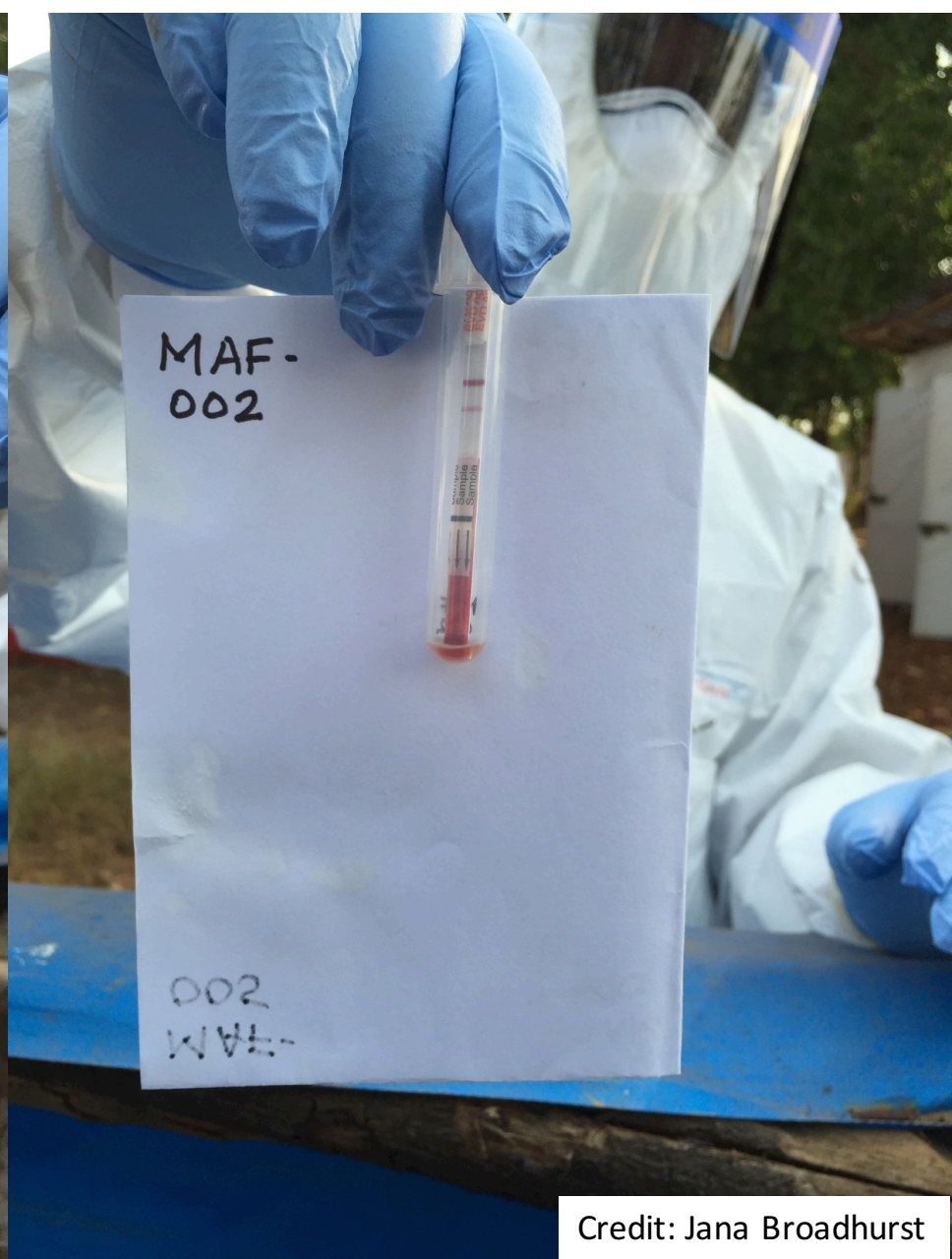
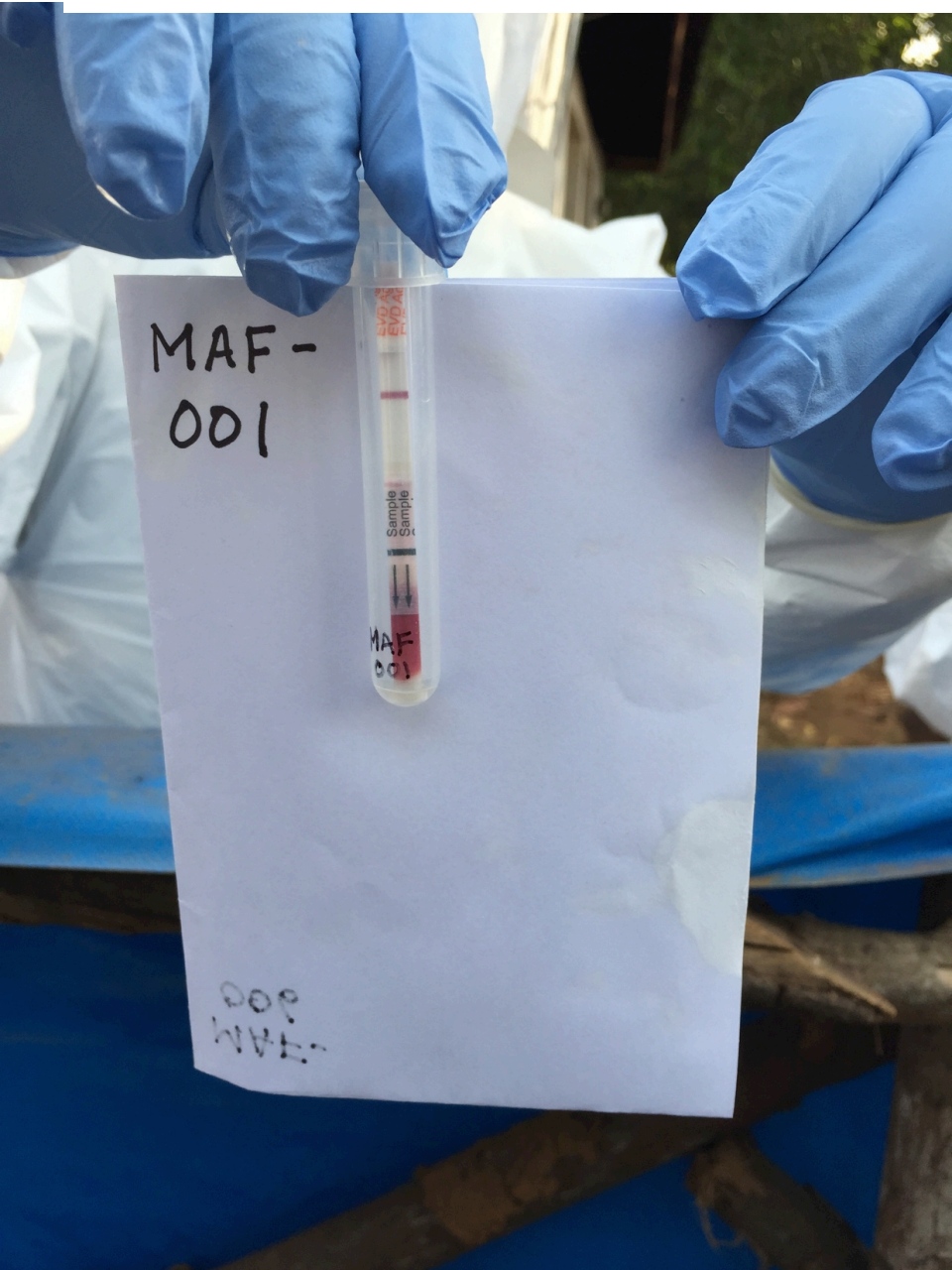
POC Evaluation

- n=106 individuals with suspected EVD in the field (Sierra Leone)
- Test: Corgenix ReEBOV Antigen Rapid Test on finger stick samples vs. realtime RT-PCR results (RealStar Filovirus Screen RT-PCR kit 1.0; altona Diagnostics GmbH) on venepuncture plasma samples

Lab Evaluation

- n=284 whole blood from individuals with suspected EVD
- Test: Corgenix ReEBOV Antigen Rapid Test vs realtime RT-PCR (Port Loko, Sierra Leone).

Alhaji Sankoh with a positive rapid diagnostic test performed on
fingerstick blood at the point of care in an Ebola care center



RT-PCR testing inside the Public Health England laboratory in Port Loko, Sierra Leone



Credit: Jana Broadhurst

	ReEBOV rapid diagnostic test on fingerstick samples (point of care; n=105)	ReEBOV rapid diagnostic test on venepuncture whole blood samples (reference laboratory; n=277)
Prevalence (% of patients/specimens that tested positive by RT-PCR)	28/105 (26.7%)	45/277 (16.2%)
Sensitivity	28/28 (100%; 87.7–100)	45/45 (100%; 92.1–100)
Specificity	71/77 (92.2%; 83.8–97.1)	214/232 (92.2%; 88.0–95.3)
Negative predictive value	71/71 (100%; 94.9–100)	214/214 (100%; 98.3–100)
Positive predictive value	28/34 (82.4%; 65.5–93.2)	45/63 (71.4%; 58.7–82.1)
Data are n/N (%) or n/N (%; 95% CI). RT-PCR=real-time reverse transcription PCR. *The altona real-time RT-PCR assay was done on fresh venepuncture plasma samples.		
Table 1: Performance of the ReEBOV rapid diagnostic test versus real-time RT-PCR (altona)*		

- Rapid+/PCR-
6/16 (38%) positive by a referee real-time RT-PCR assay (Trombley assay)
- Rapid-/PCR-
3/18 (17%) positive by Trombley

GeneXpert Ebola PCR Assay



Feasibility of Xpert Ebola Assay in Médecins Sans Frontières Ebola Program, Guinea

Rafael Van den Bergh, Pascale Chaillet, Mamadou Saliou Sow, Mathieu Amand, Charlotte van Vyve,
Sylvie Jonckheere, Rosa Crestani, Armand Sprecher, Michel Van Herp, Arlene Chua,
Erwan Piriou, Lamine Koivogui, Annick Antierens

Médecins Sans Frontières

➤ Lab Evaluation

Tested 218 whole blood from patients with suspected EVD, on treatment, and convalescence
Compared Xpert to lab-developed RT-PCR

Sample type	No. positive by Xpert Ebola Assay	No. (%) positive by routine PCR
Total	26	18 (69)
Diagnosis 1	8	8 (100)
Diagnosis 2	0	0
After transfusion	12	9 (75)
Convalescent phase	6	1 (17)

Feasibility of Xpert Ebola Assay in Médecins Sans Frontières Ebola Program, Guinea

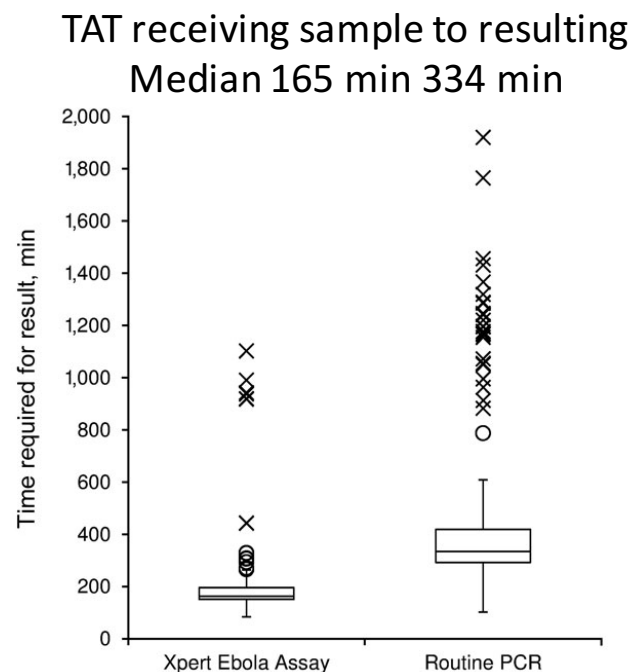
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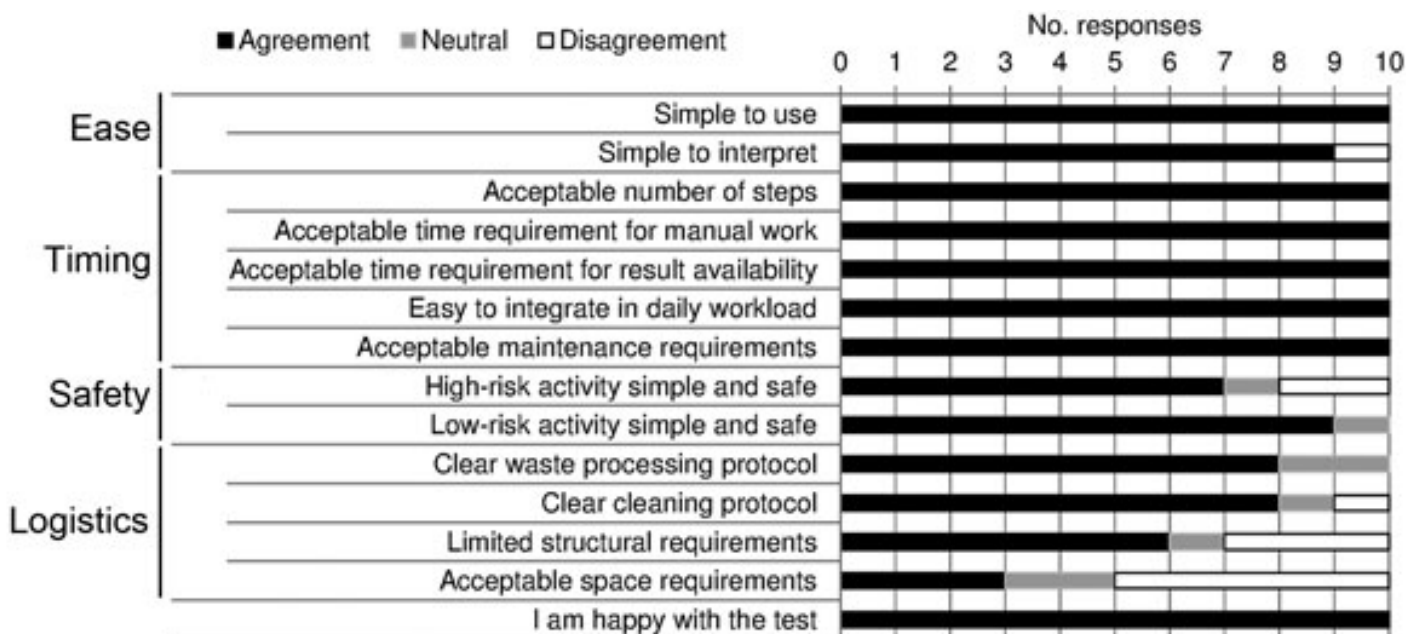
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LB-1. Field Laboratory Evaluation of the GeneXpert Ebola Assay for Diagnosis of Ebola Virus Disease in Sierra Leone

Amanda Semper, Jana Broadhurst, Jade Richards, Geraldine M. Foster, Andrew Simpson, Elisabetta Groppelli, Julie Johnson, J. Dan Kelly, Tim Brooks, Christopher H. Logue, Ann Miller, Megan B. Murray, and Nira R. Pollock, MD, PhD

Evaluation at a field biocontainment laboratory in Sierra Leone

- Individuals with suspected EVD
- Specimens: Whole blood (n = 218) and buccal swab (n = 71) specimens
- Tests: Xpert vs routine EVD testing by RT-PCR (“Trombley assay”)
 - Compared whole blood sampling methods (pipette vs swab) on 84/218

Loading up the Cepheid GeneXpert for delivery to the Public Health England field laboratory in Port Loko in preparation for the field validation study



Credit: Jana Broadhurst

Bringing the GeneXpert into the PHE field lab in Port Loko



Credit: Jana Broadhurst



LB-1. Field Laboratory Evaluation of the GeneXpert Ebola Assay for Diagnosis of Ebola Virus Disease in Sierra Leone

➤ Invalid Xpert results

7/218 (3.2%) whole blood and 7/71 (9.9%) buccal swab

➤ Tromboley PCR+

On whole blood: 22/22 Xpert+ (Sensitivity 100%)

On buccal swab: 20/20 Xpert+ (Sensitivity 100%)

➤ Tromboley PCR-

On whole blood: 181/189 Xpert- (Specificity 95.8%) → 7/8 from previously PCR+ →
Xpert Specificity 99.5%

On buccal swab 44/44 Xpert- (Specificity 100%)

➤ Whole blood sampled with pipette vs Swab concordant for 78/79 (98.7%)

Sample reagent reduced EBOV titer by ≥ 6 logs (Pinsky et al PLoS One 2015)

Brazil state announces Zika fever emergency

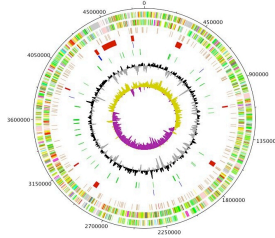
🕒 1 December 2015 | [Latin America & Caribbean](#)



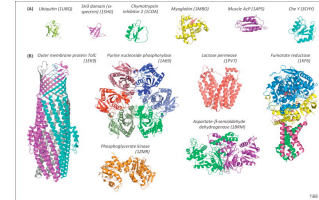
Science Photo Library

Aedes aegypti mosquito is found throughout tropical Africa and in parts of South America

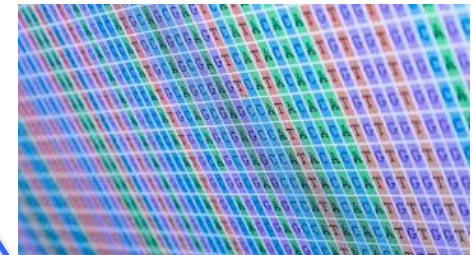
Recipe for POC Diagnostic Development



Genomics/
Proteomics Databases



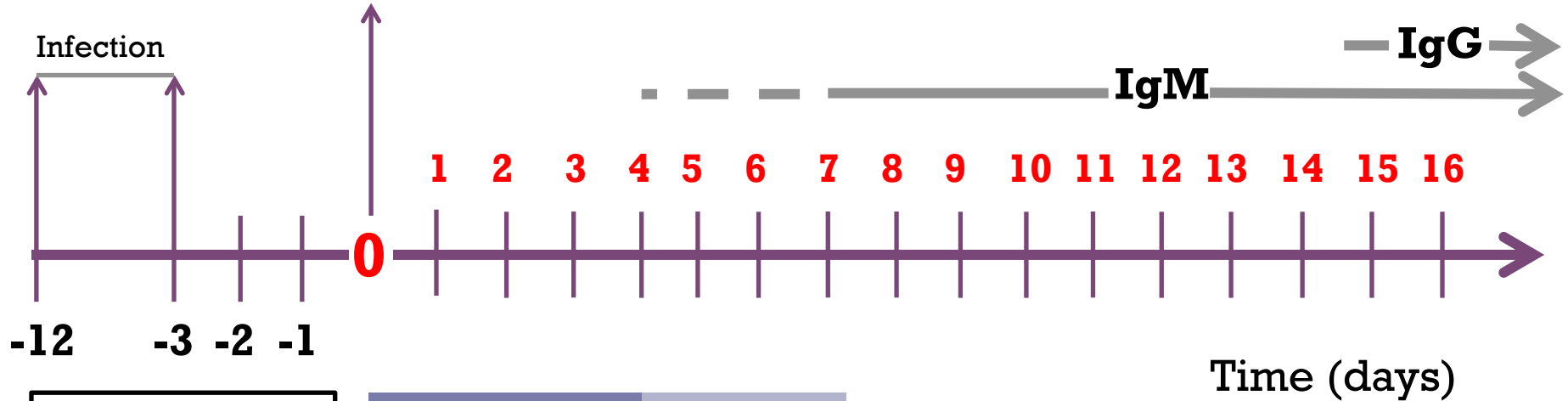
Molecular
Diagnostic
Platforms



Bioinformatics
Tools

Zika Virus Diagnostics

First Symptoms



Incubation
Period

Viral RNA

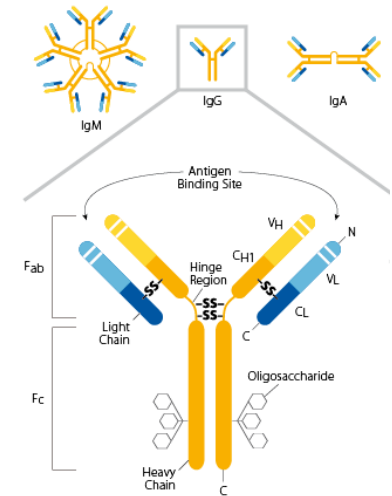
PCR Diagnosis

Serological Diagnosis

Zika Virus Diagnostics: Serology

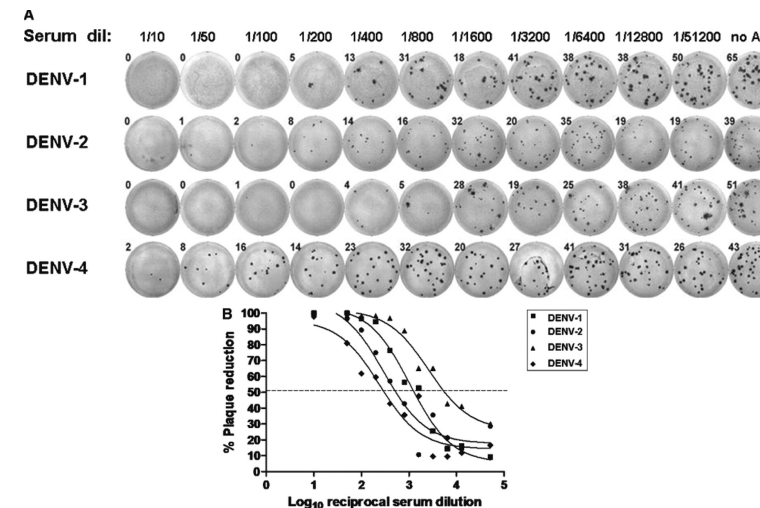
• ZIKV IgM Antibodies

- CDC IgM ELISA: extensive flavivirus cross reaction.
- Previous Infection or vaccination with other flaviviruses (*e.g.*, DENV, JEV, YFV) may result in false positive results.
- Note: returning travelers frequently have an initial diagnosis of dengue, due to false-positive DENV IgM tests.
- CDC IgM ELISA received FDA EUA
- Euroimmun offers CE marked Zika virus ELISA assays



• Plaque Reduction Neutralization Testing (PRNT)

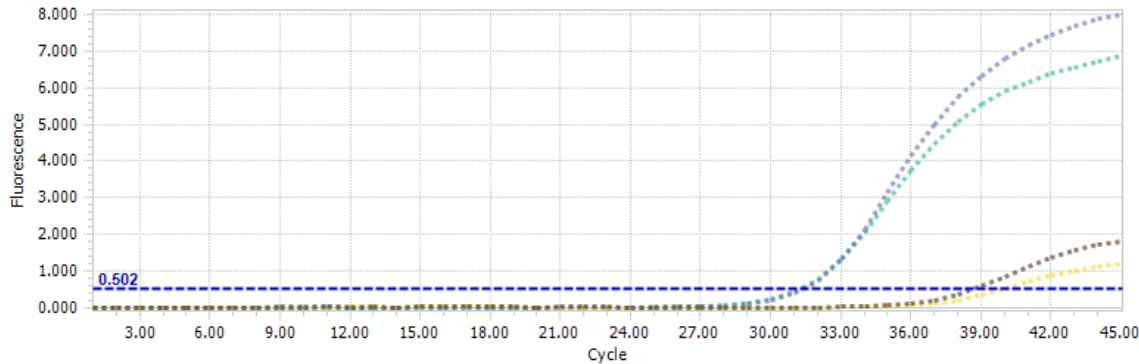
- PRNT90 may help distinguish ZIKV infection from infection with DENV or other flaviviruses.
- ≥ 4 -fold rise in titer in acute and convalescent sera or ≥ 4 -fold greater ZIKV titer compared to other flaviviruses in a single specimen.



• NS1 Antigen Testing

- Not currently available

Zika Virus Diagnostics: Nucleic Acid Amplification Testing



- RNA Detection
 - Serum: short duration of RNAemia, lower viral loads compared to DENV or CHIKV.
 - Saliva: higher rate of detection, duration similar to serum.
 - Urine: may be detectable 7 or more days after undetectable in serum.
- ZIKV CDC real-time, reverse transcriptase-PCR (rRT-PCR)
 - Most commonly performed globally.
 - Designed based on sequences from the 2007 Yap Islands outbreak.
 - Lanciotti *et al.*, Emerging Infectious Diseases 2008
- Other rRT-PCR Options
 - Altona Diagnostics offers a CE marked rRT-PCR assay.
 - Several laboratory developed ZIKV rRT-PCR assays have been published.

Stanford Zika Virus Testing



		CDC ZIKV rRT-PCR		
		Positive	Negative	Total
Stanford Multiplex ZIKV	Positive	25	31*	56
	Negative	1	76	77
	Total	26	107	133

- Stanford Multiplex ZIKV/CHIKV/DENV rRT-PCR
 - Detects and distinguishes between these three co-circulating arboviruses.
 - ZIKV was detected in significantly more Nicaraguan samples using the Stanford assay than the CDC ZIKV rRT-PCR ($p < 0.001$).
 - In a larger set of samples from patients with suspected arboviral infection
 - 73.7% (255/346) tested positive.
 - Co-infections accounted for 20.1% (53/255) of the positives: DENV-CHIKV ($n=36$), CHIKV-ZIKV ($n=10$), DENV-ZIKV ($n=3$), and DENV-CHIKV-ZIKV ($n=4$).

Overview

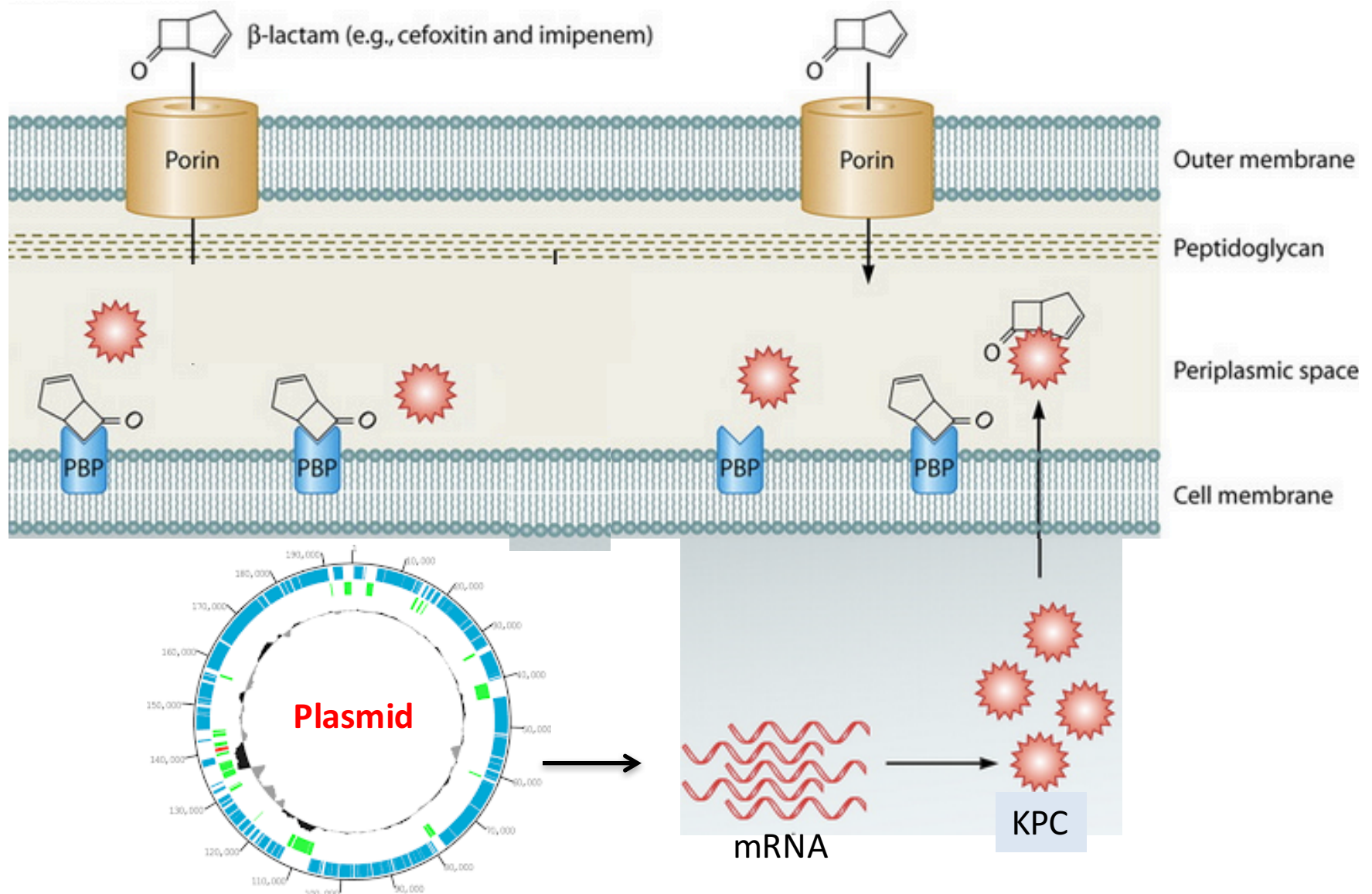
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Carbapenem-Resistant Enterobacteriaceae (CRE)

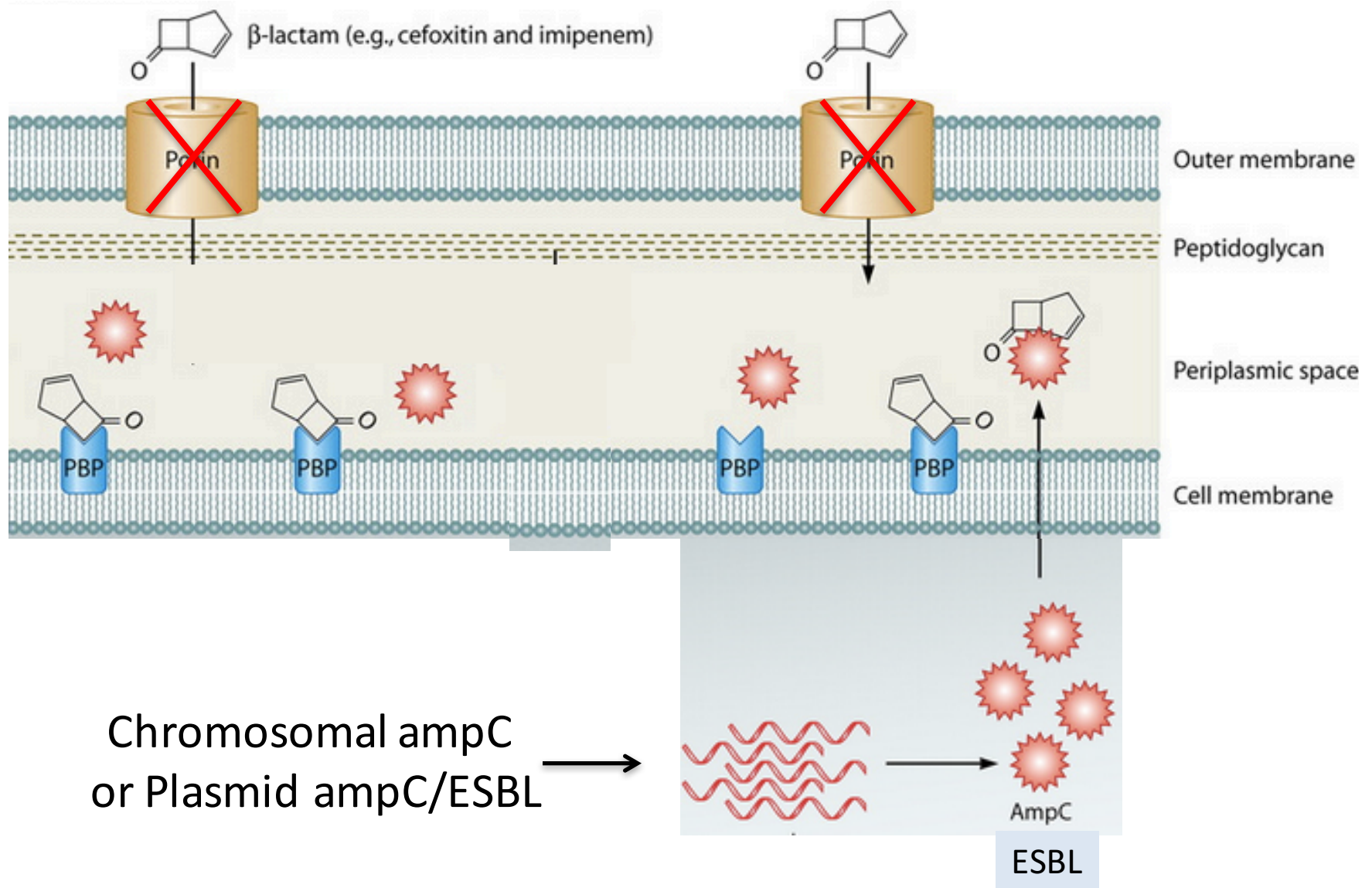


2015 CDC definition: Resistant to imipenem, meropenem, doripenem or ertapenem

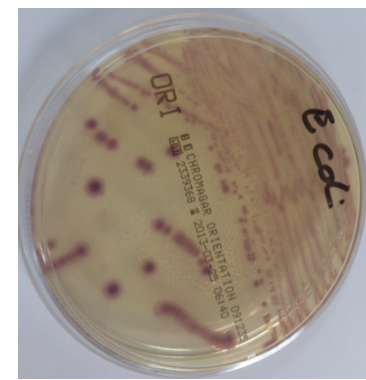
Mechanism of Carbapenem Resistance



Mechanism of Carbapenem Resistance



Indian man with pyelonephritis.
 Visiting daughter in Silicon Valley.
 Ureteral stent placed in India for kidney stones.
 Urine culture: *E. coli* >100,000/mL

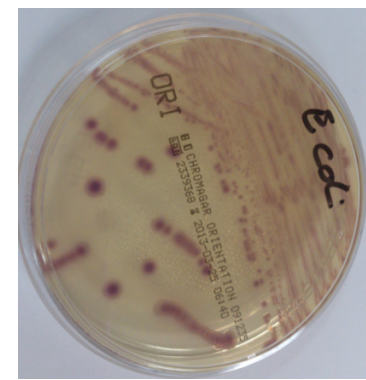


Susceptibility

	MIC by Vitek 2	
Amikacin	>=64 ug/mL ...	RESISTANT
Amoxicillin/Clavulanic Acid	>=32 ug/mL ...	RESISTANT
Ampicillin	>=32 ug/mL ...	RESISTANT
Cefazolin	>=64 ug/mL ...	RESISTANT ¹
Cefepime	>=64 ug/mL ...	RESISTANT
Cefoxitin	>=64 ug/mL ...	RESISTANT
Ceftazidime	>=64 ug/mL ...	RESISTANT
Ceftriaxone	>=64 ug/mL ...	RESISTANT
Ciprofloxacin	>=4 ug/mL (...)	RESISTANT
Doxycycline		
Ertapenem	>=8 ug/mL (...)	RESISTANT
Gentamicin	>=16 ug/mL ...	RESISTANT
Imipenem		
Levofloxacin	>=8 ug/mL (...)	RESISTANT
Meropenem	8 ug/mL (MIC)	RESISTANT
Nitrofurantoin	64 ug/mL (MIC)	INTERMEDIATE
Piperacillin/Tazobactam	>=128 ug/mL...	RESISTANT
Tetracycline	>=16 ug/mL ...	RESISTANT
Tobramycin	>=16 ug/mL ...	RESISTANT
Trimethoprim/Sulfamethoxazole.	>=320 ug/mL...	RESISTANT

The image shows a petri dish containing a yellowish agar medium. Three white antibiotic disks are placed on the surface, each surrounded by a red circle indicating the zone of inhibition. The disks are labeled "E", "T", and "P". A vertical ruler scale is visible on the right side of the dish, marked from 0.002 to 32 mm. Handwritten text on the agar includes "ETP", "Klebsiella pneumoniae", and "ATCC 35967".

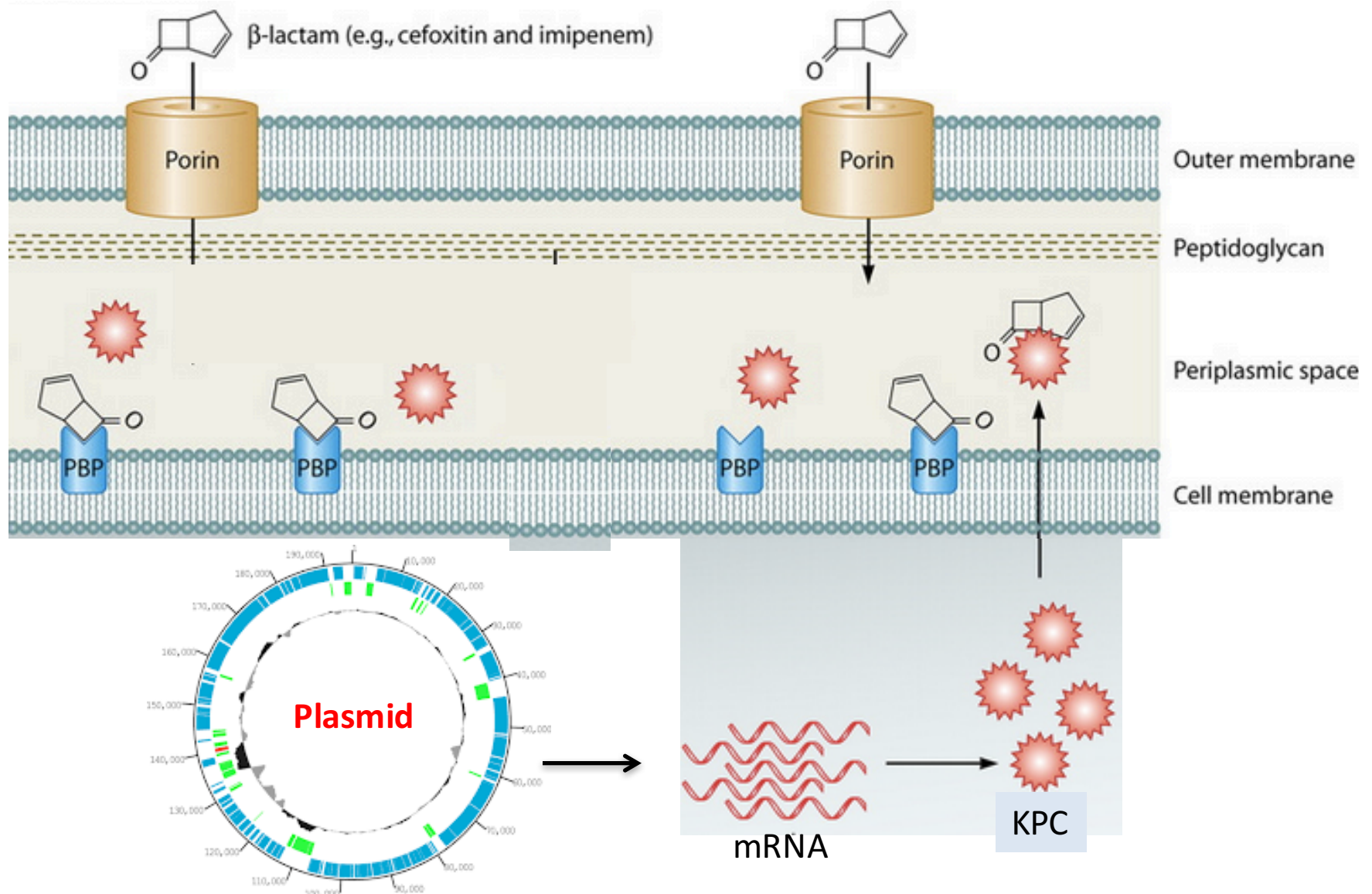
Indian man with pyelonephritis.
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 Ureteral stent placed in India for kidney stones.
 Urine culture: *E. coli* >100,000/mL



Susceptibility

	MIC by Vitek 2		Disk Diffusion	MIC by Etest
Amikacin	>=64 ug/mL ...	RESISTANT		
Amoxicillin/Clavulanic Acid	>=32 ug/mL ...	RESISTANT		
Ampicillin	>=32 ug/mL ...	RESISTANT		
Cefazolin	>=64 ug/mL ...	RESISTANT ¹		
Cefepime	>=64 ug/mL ...	RESISTANT		
Cefoxitin	>=64 ug/mL ...	RESISTANT		
Ceftazidime	>=64 ug/mL ...	RESISTANT		
Ceftolozane/Tazobactam				>256 ug/mL ... R
Ceftriaxone	>=64 ug/mL ...	RESISTANT		
Ciprofloxacin	>=4 ug/mL (...)	RESISTANT		
Doxycycline				>256 ug/mL ... R
Ertapenem	>=8 ug/mL (...)	RESISTANT	RESISTANT	
FOSFOMYCIN			SUSCEPTIBLE	
Gentamicin	>=16 ug/mL ...	RESISTANT		
Imipenem			RESISTANT	
Levofloxacin	>=8 ug/mL (...)	RESISTANT		
Meropenem	8 ug/mL (MIC)	RESISTANT	RESISTANT	
Nitrofurantoin	64 ug/mL (MIC)	INTERMEDIATE		
Piperacillin/Tazobactam	>=128 ug/mL...	RESISTANT		
Polymixin B				1 ug/mL (MIC) No interpret
Tetracycline	>=16 ug/mL ...	RESISTANT		
Tobramycin	>=16 ug/mL ...	RESISTANT		
Trimethoprim/Sulfamethoxazole.	>=320 ug/mL...	RESISTANT		

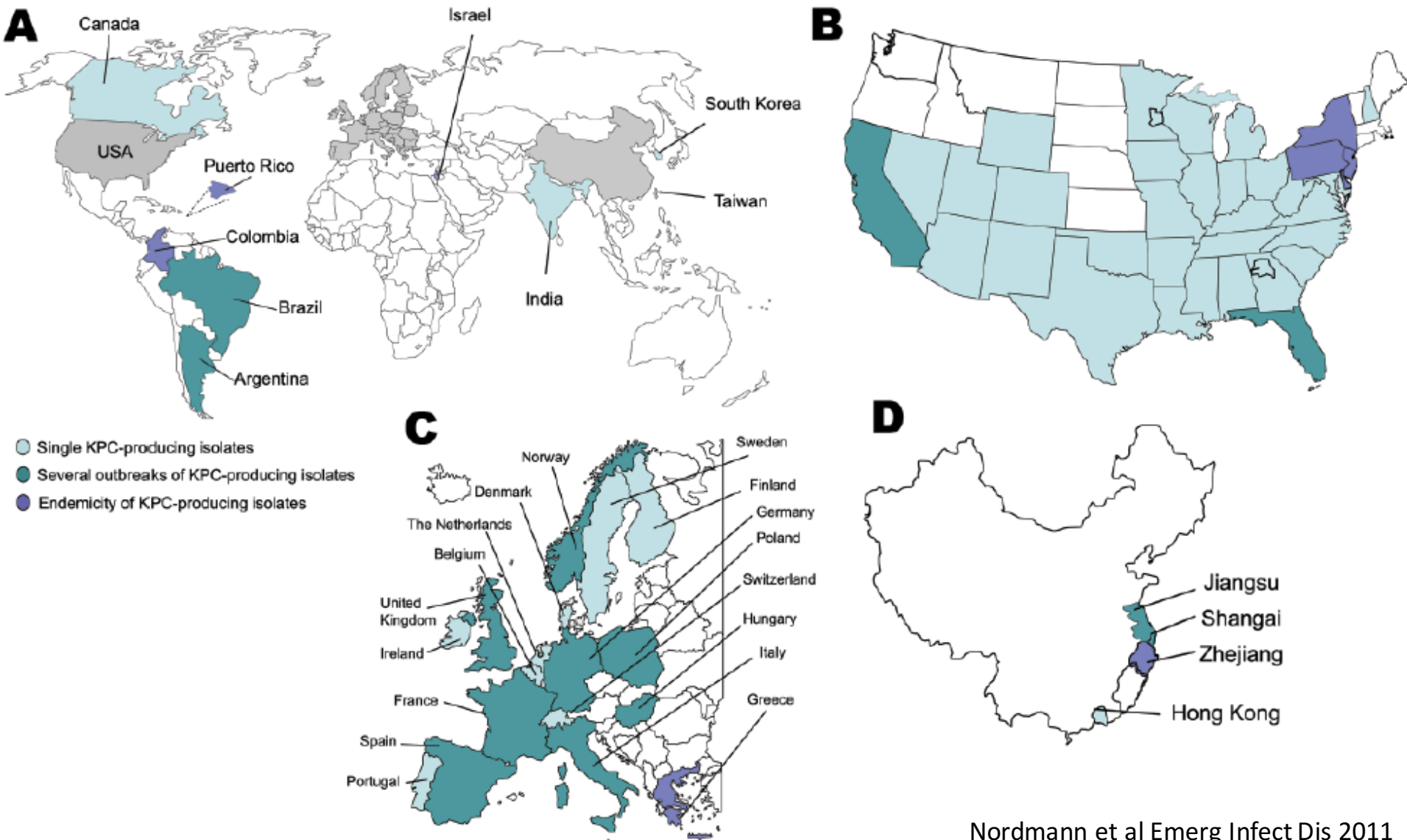
Mechanism of Carbapenem Resistance



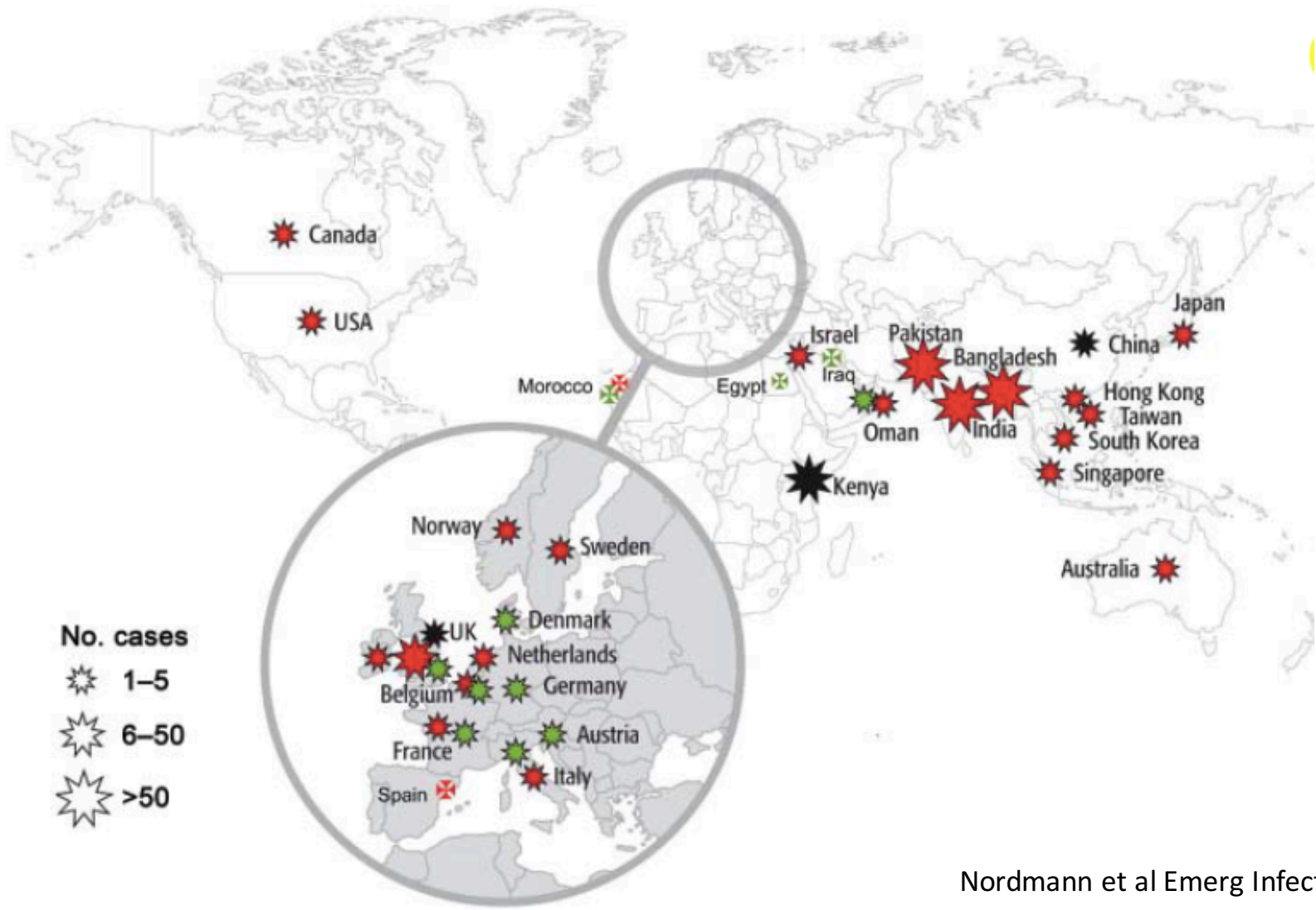
Emergence of Carbapenemases

Enzyme	Molecular Class	Location	Year of Isolation	City,Country of origin
IMP	B	Plasmid	1991	Japan
KPC	A	Plasmid	1996	NC, USA
VIM	B	Plasmid	1996	Verona, Italy
OXA-48-like	D	Plasmid	2001	Turkey
NDM	B	Plasmid	2008	India

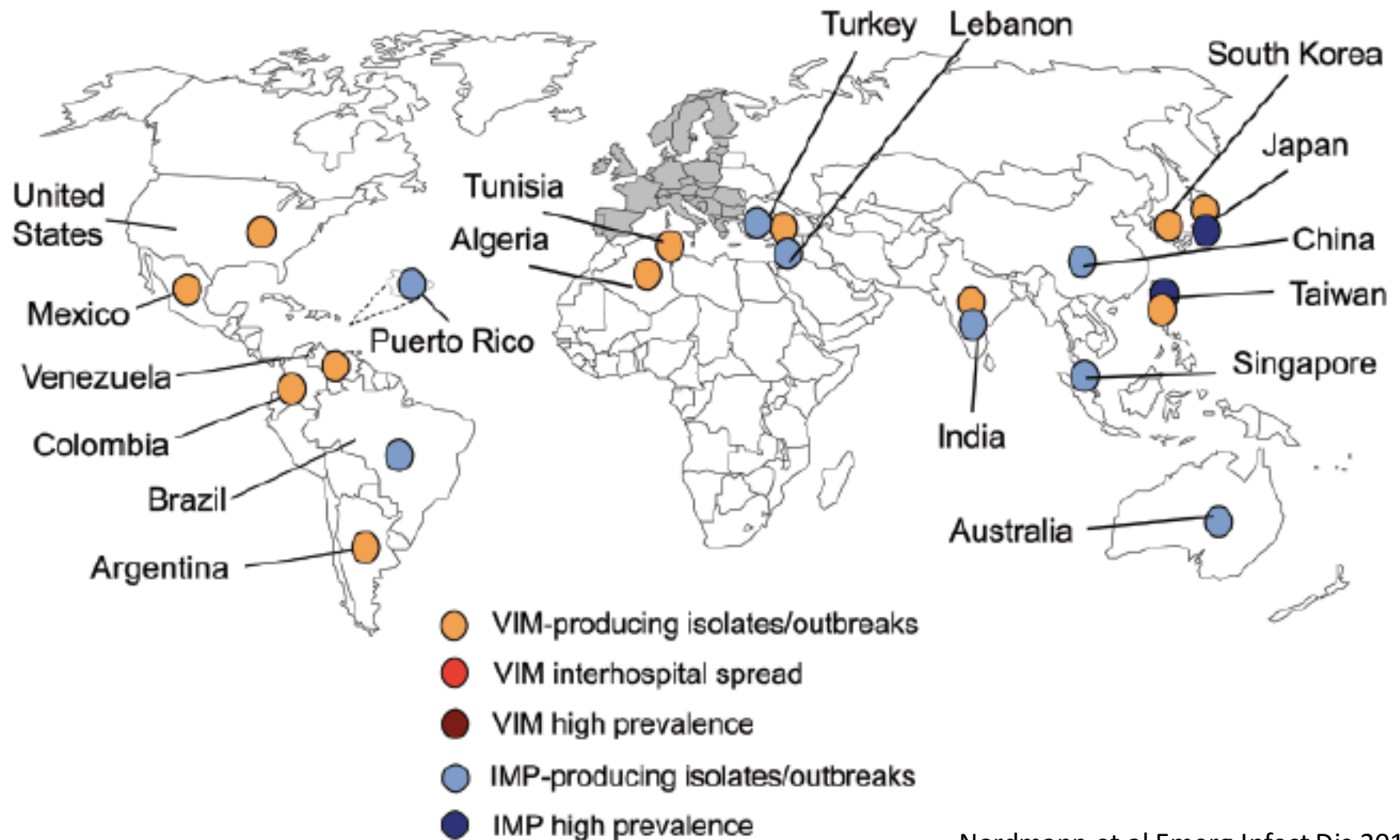
Geographic Distribution of *Klebsiella pneumoniae* Carbapenemase (KPC) producers



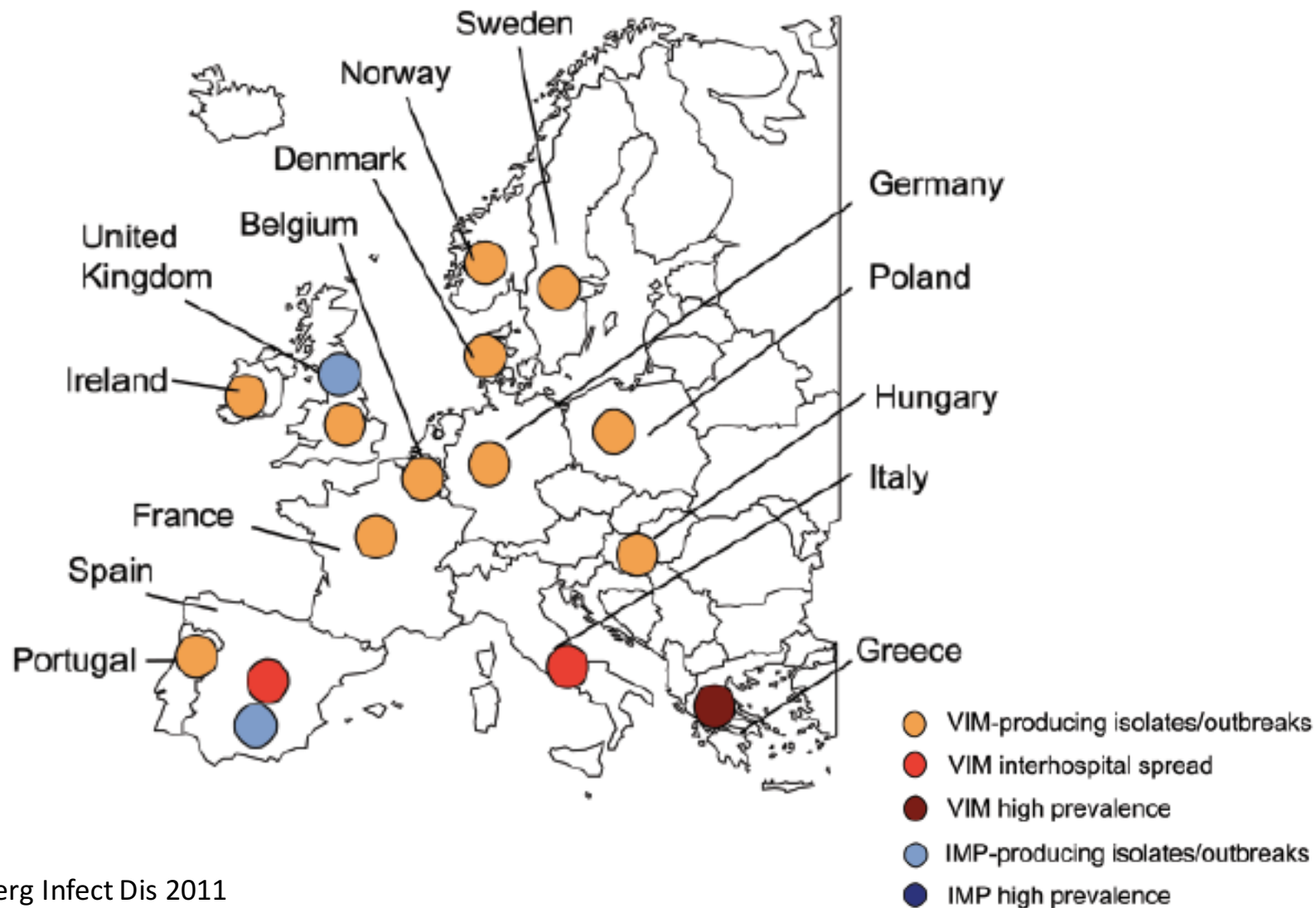
Geographic Distribution of New Delhi Metallo- β -Lactamase Producers



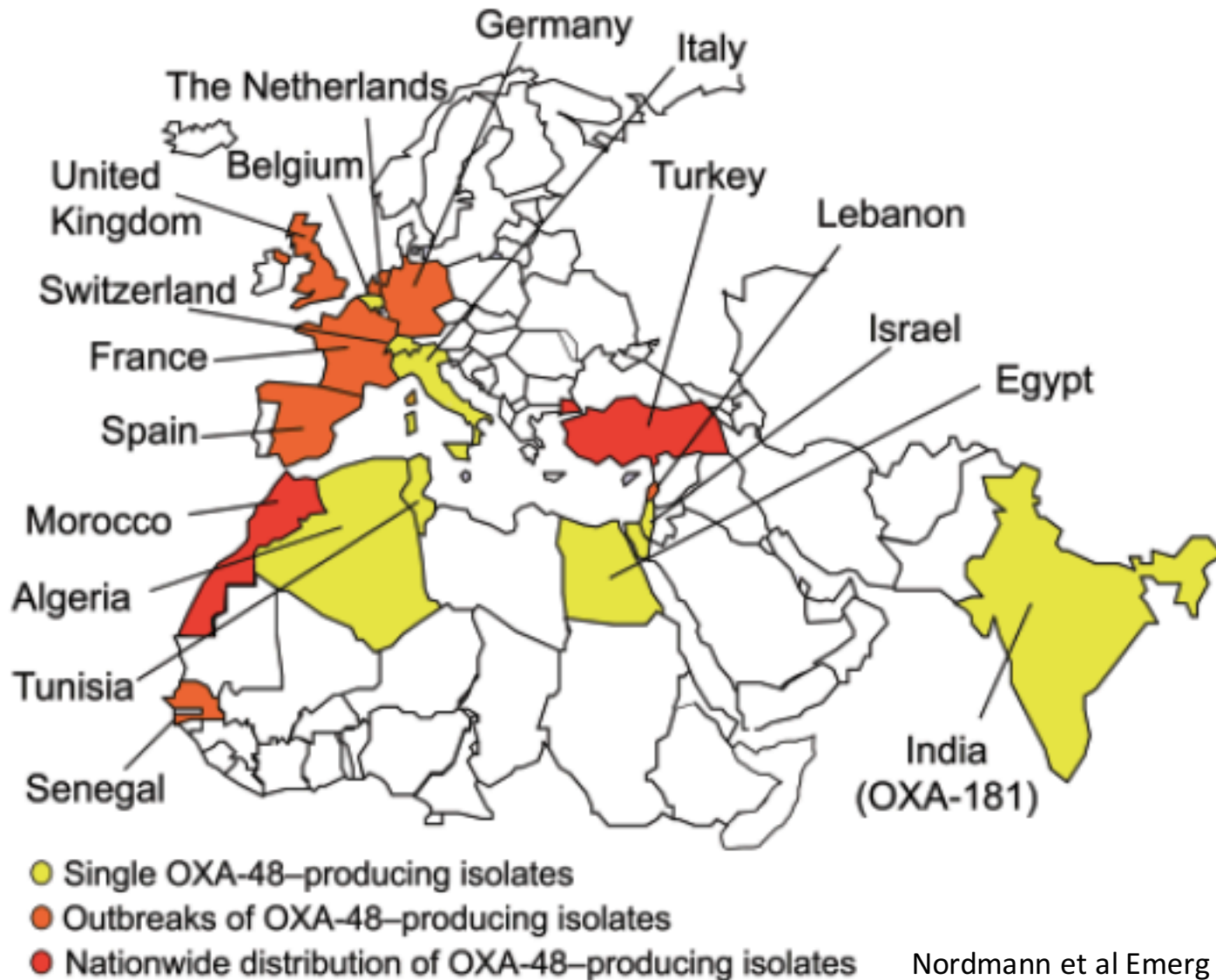
Distribution of Verona Integron-Encoded Metallo- β -Lactamase (VIM) & IMP Producers



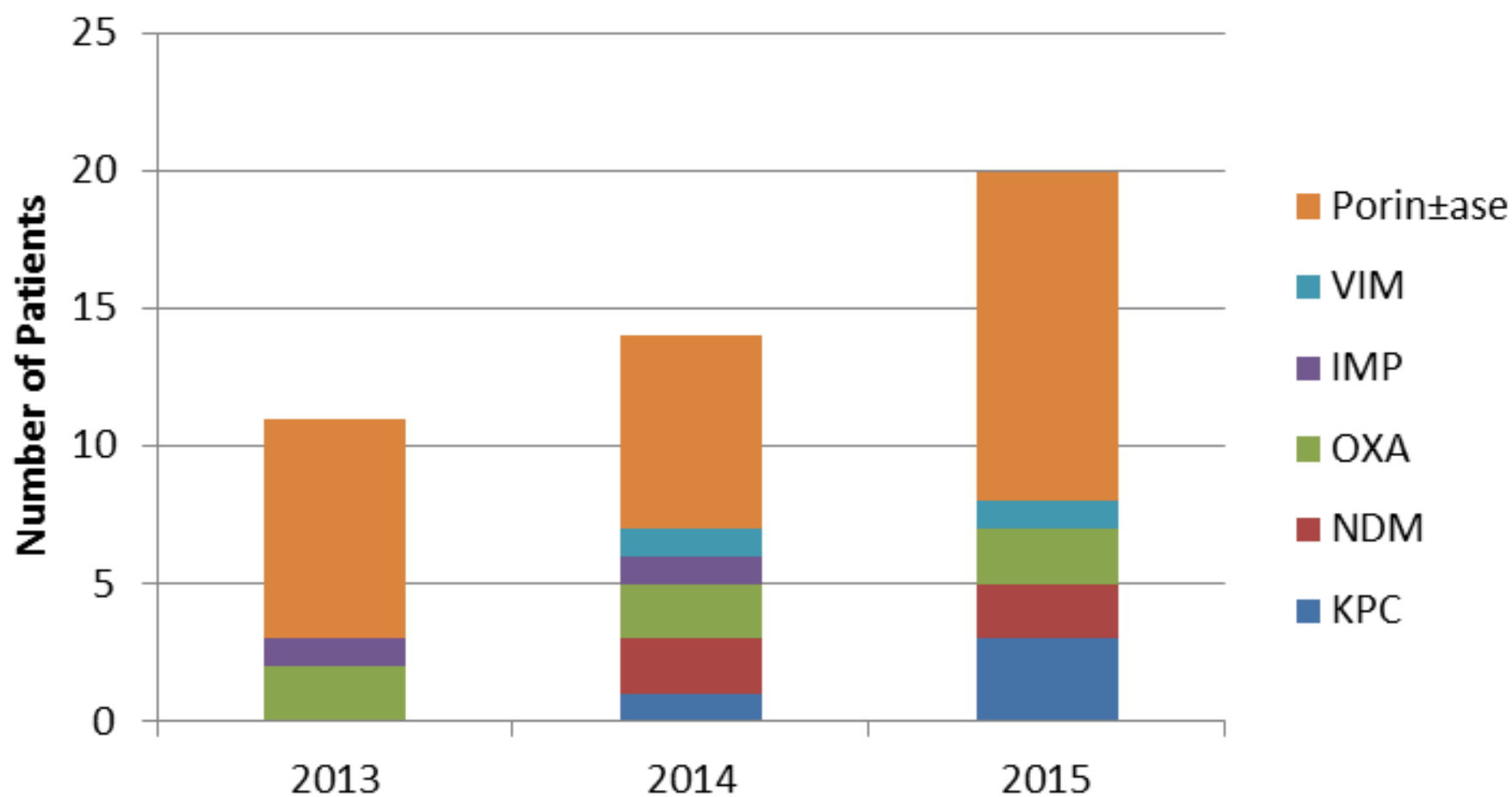
Distribution of Verona Integron-Encoded Metallo- β -Lactamase (VIM) & IMP Producers



Geographic Distribution of Oxacillinase-48-like (OXA-48-like) Type Producers

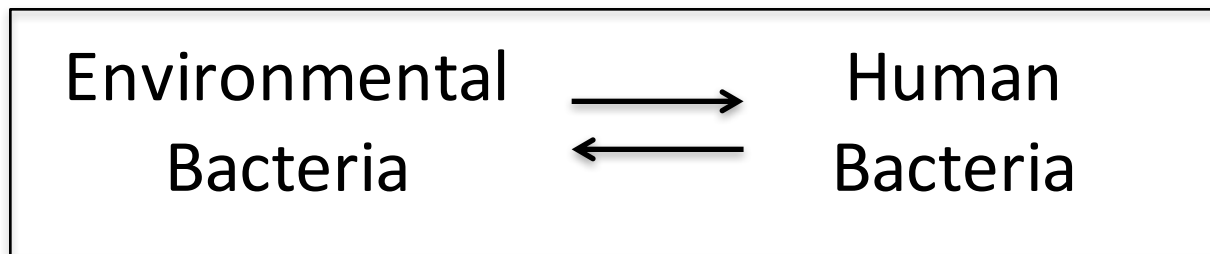


Mechanism of CRE at Stanford Hospitals

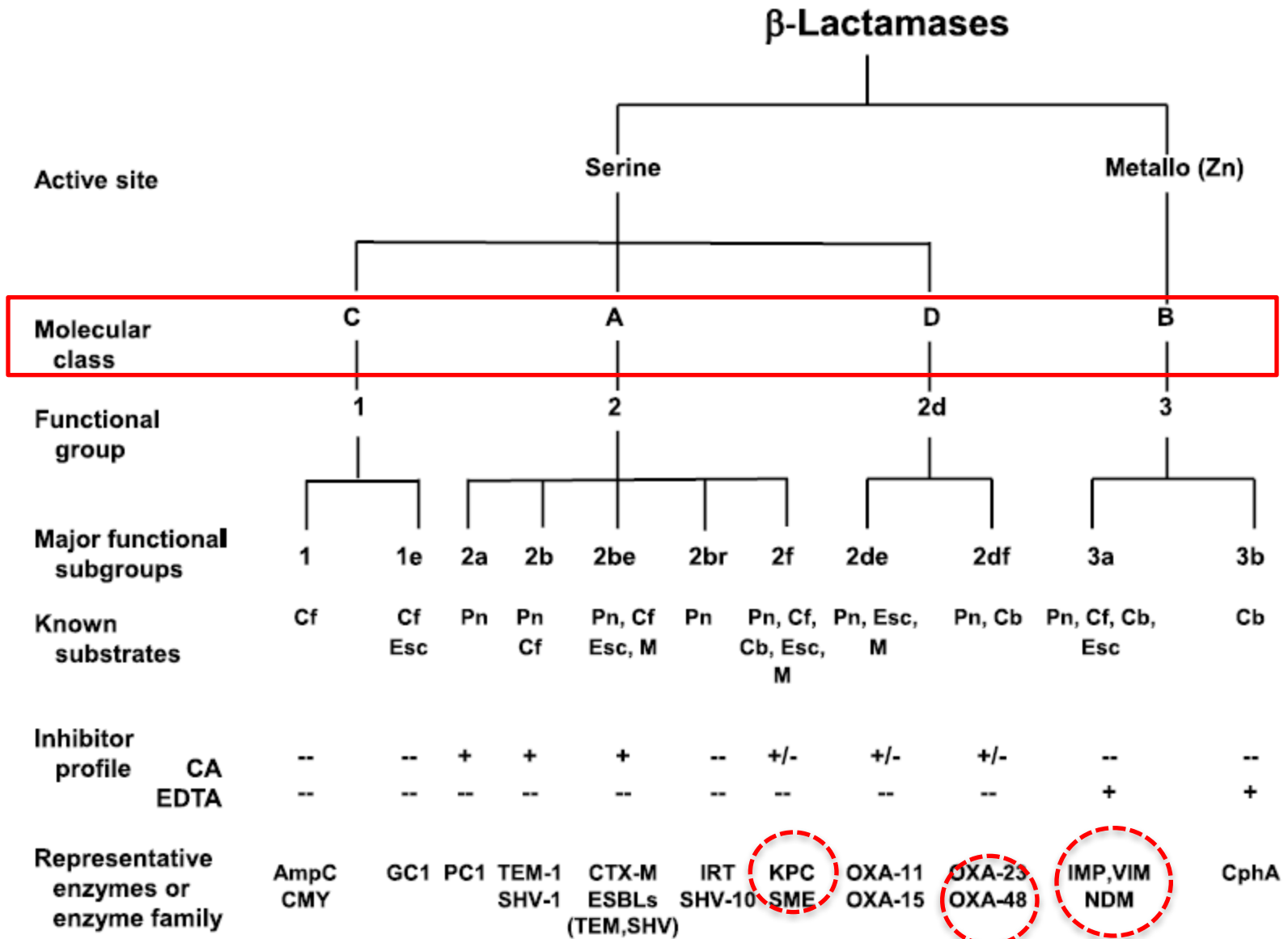


Origins of Carbapenemases

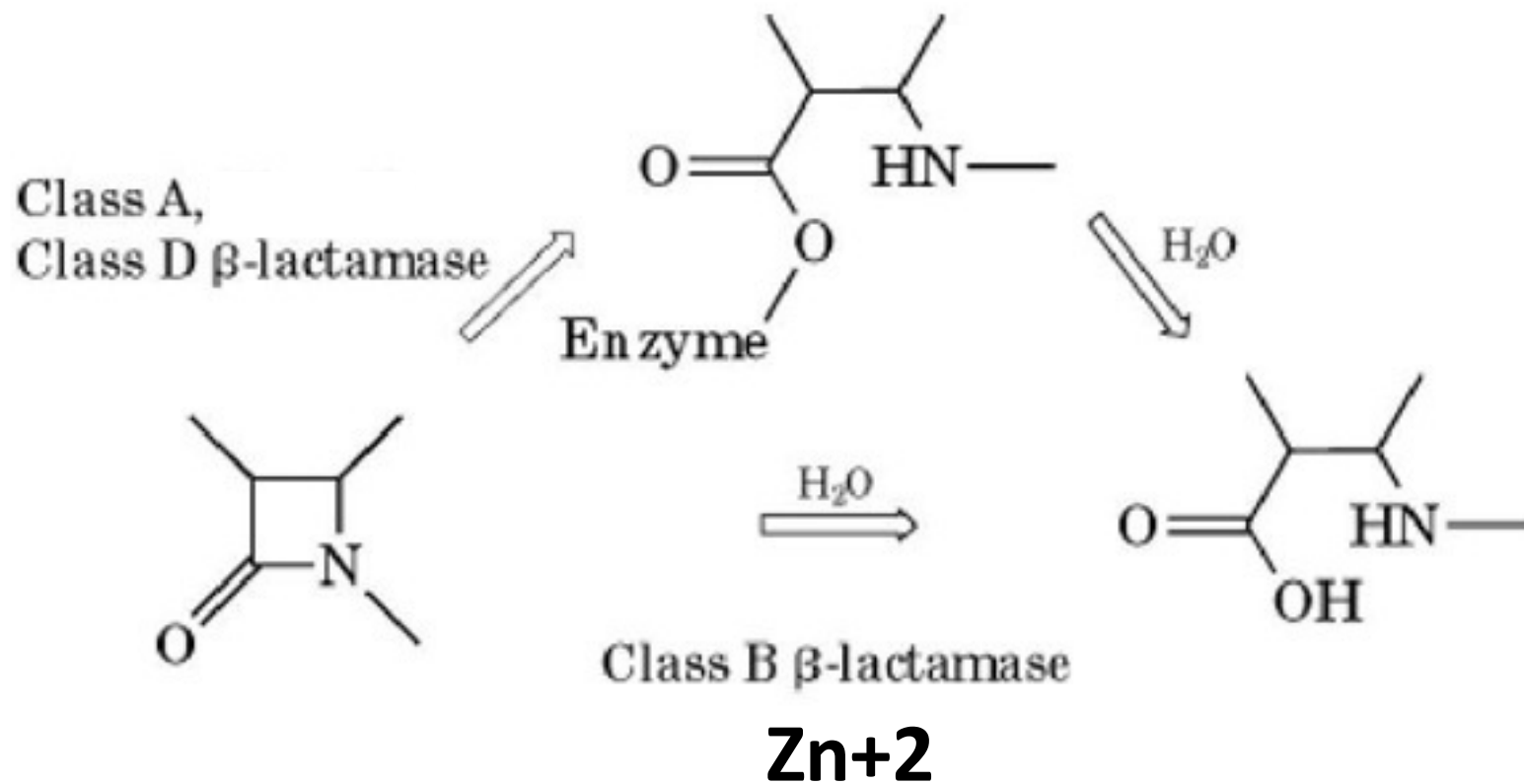
- Carbapenem design inspired by thienamycin, produced by the soil organism *Streptomyces cattleya*
- Carbapenemases originate from environmental organisms: OXA-48-like from *Shewanella* spp.; NDM from *Pseudoxanthomonas*



Carbapenemase Classification

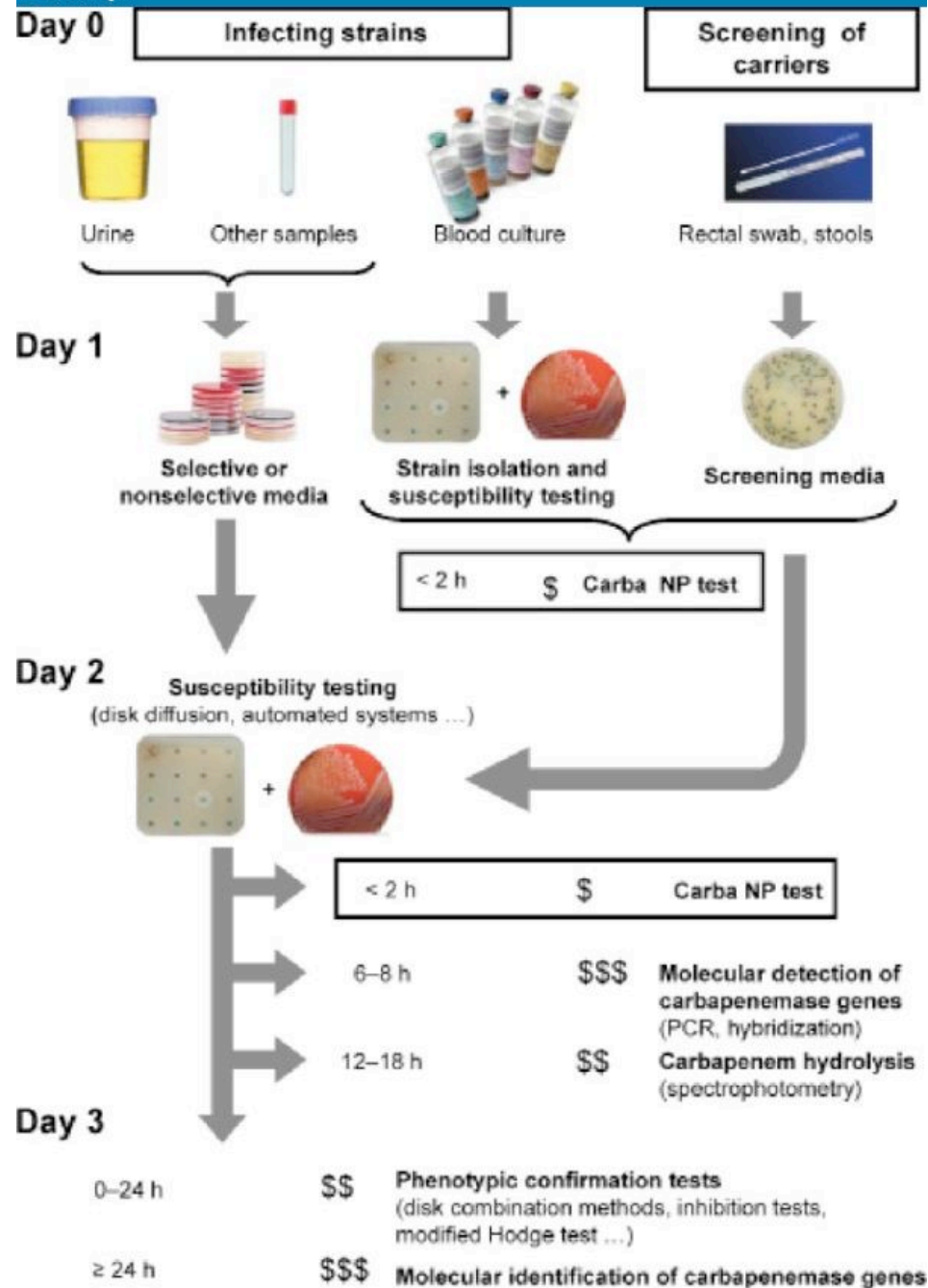


Carbapenemase Mechanisms



Laboratory Diagnostics for CRE

	Method	Need Culture	Duration (hrs)	Cost	Reveals Mechanism
Phenotypic	ChromAgar	Yes	24	\$	No
	Carba NP	Yes	0.5-2	\$	No
	MALDI	Yes	0.5	\$	No
Genotypic	Xpert PCR	Optional	1	\$\$	Yes
	LDT PCR	Optional	2	\$	Yes



Culture Based Detection of CREs

HardyCHROM™ CRE Agar

Organism	Description	Photo	Color
<i>Klebsiella</i> , <i>Enterobacter</i> , and <i>Serratia</i> spp.	Dark blue colonies		
<i>Escherichia coli</i>	Pink colonies with dark pink centers		

The main problem remains detection of OXA-48 producers that are susceptible to cephalosporins and have low-level resistance to carbapenems when not co-producing an ESBL



Significant Variations in the Level of Resistance to Carbapenems for Each Enzyme

Carbapenemase	MIC, mg/L		
	Imipenem	Meropenem	Ertapenem
KPC	0.5→64	1→64	0.5→64
Metallo β -lactamases†	0.5→64	0.25→64	0.5→64
OXA-48 type	1→64	0.5→64	0.25→64

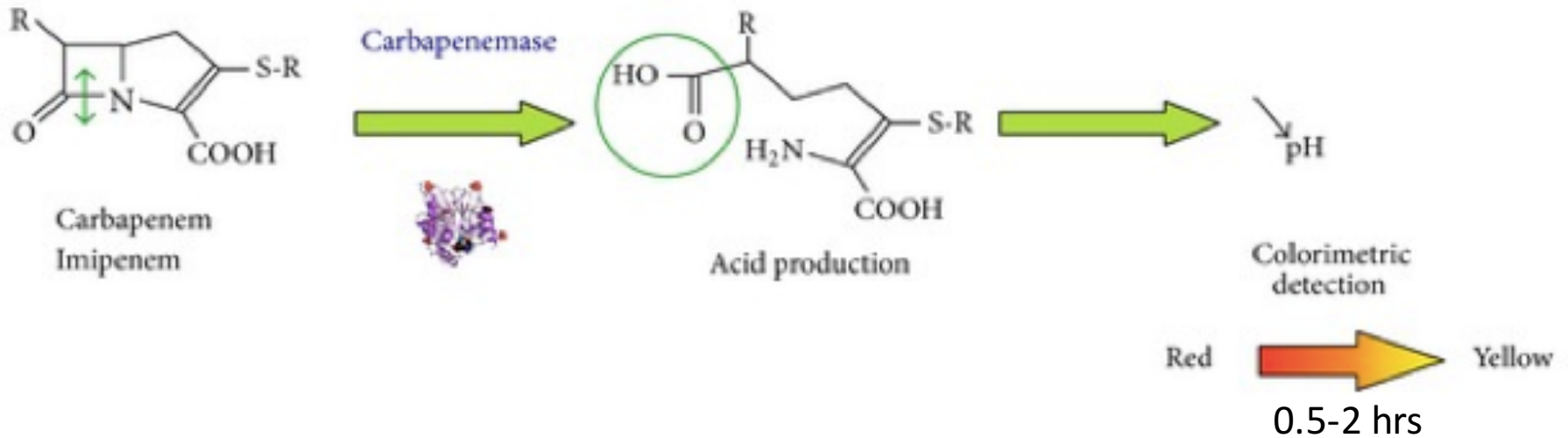
*KPC, *Klebsiella pneumoniae* carbapenemase; OXA-48, oxacillinase-48.

†Including New Delhi metallo- β -lactamase-1.

Carbapenem and oxyimino- β -lactam MICs are commonly higher for species with derepressed production of their chromosomally encoded AmpCs

Phenotypic Detection of Carbapenemases:

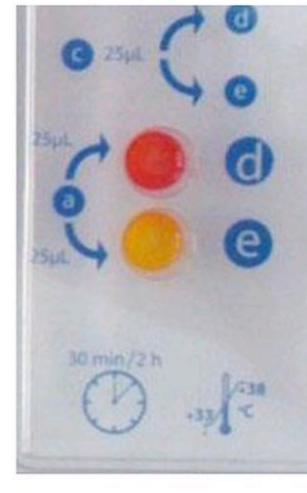
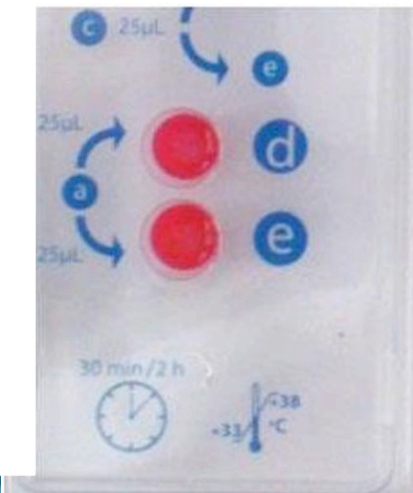
Carba NP (Nordmann et al Emerg Infect Dis 2012)



RAPIDEC® CARBA NP (bioMérieux)

First reading after 30 min

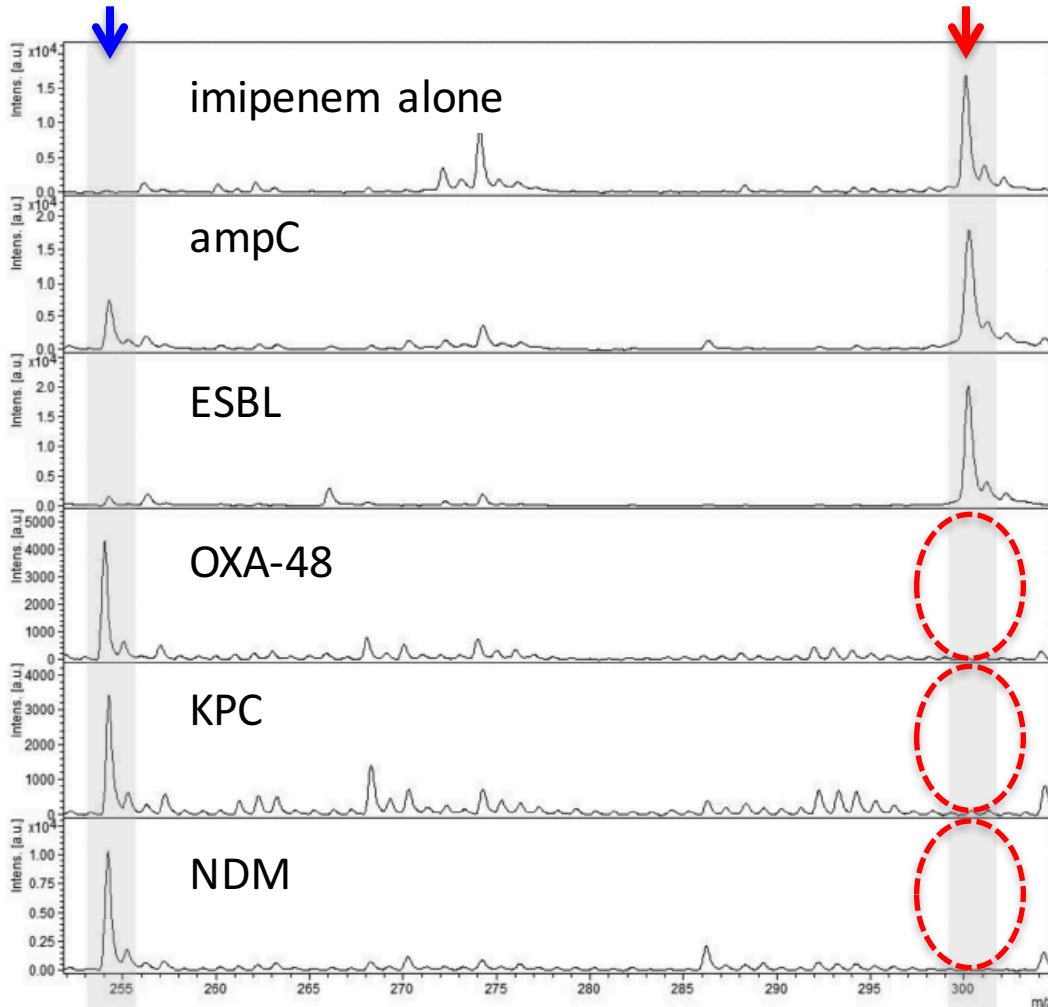
Sensitivity: 100%
Specificity: 100%
Vasoo et al JCM 2013



Phenotypic Detection of Carbapenemases: MALDI-TOF Mas Spectrometry

metabolite (254 Da m/z)

imipenem: 300 Da m/z



MALDI-TOF spectra of imipenem hydrolysis assays after a 20-min incubation at 37°C

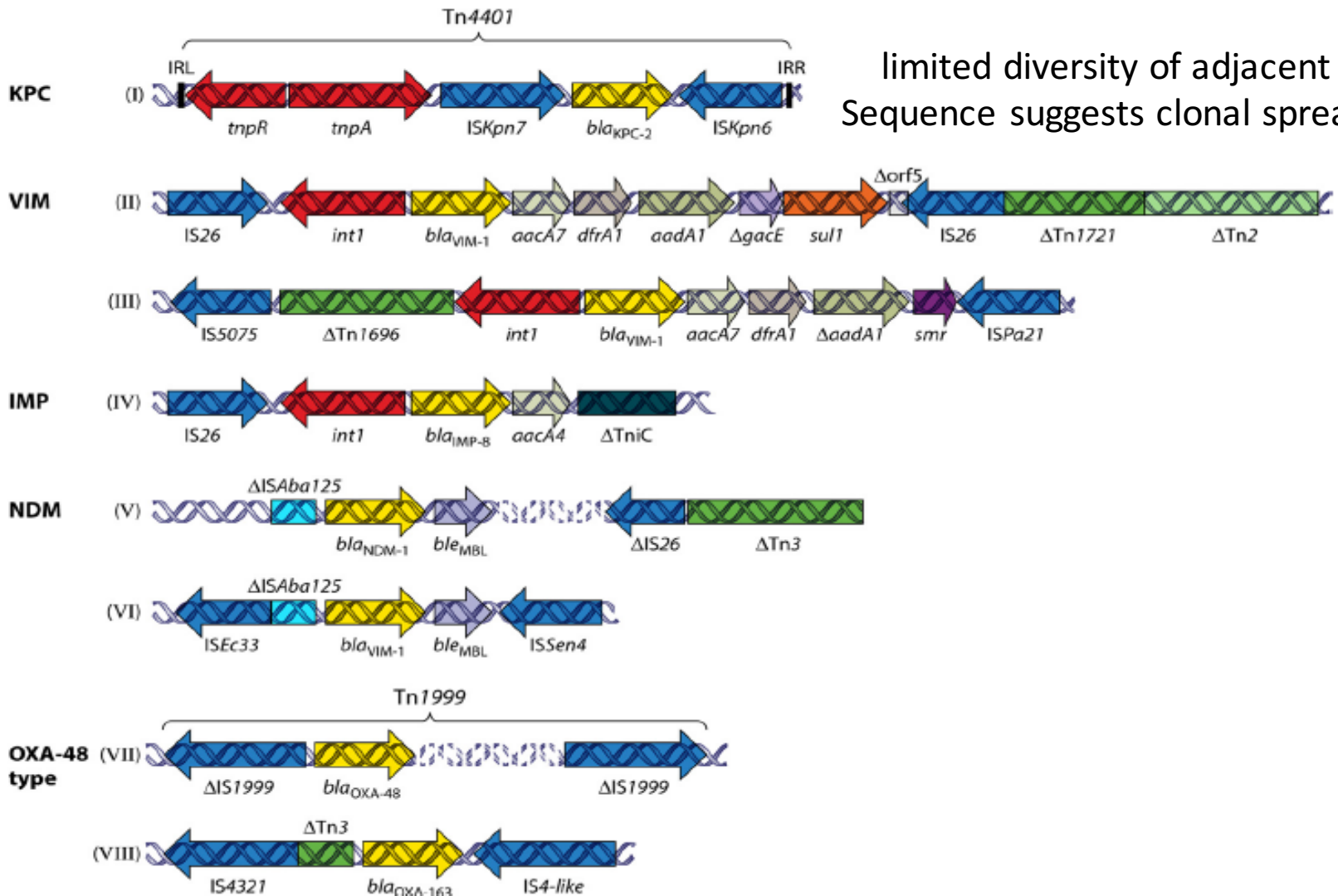
$$\frac{\text{metabolite}}{(\text{metabolite} + \text{imipenem})} \geq 0.82$$

Training set: 77 CPE & 146 non-CPE

Sensitivity: 97.8%

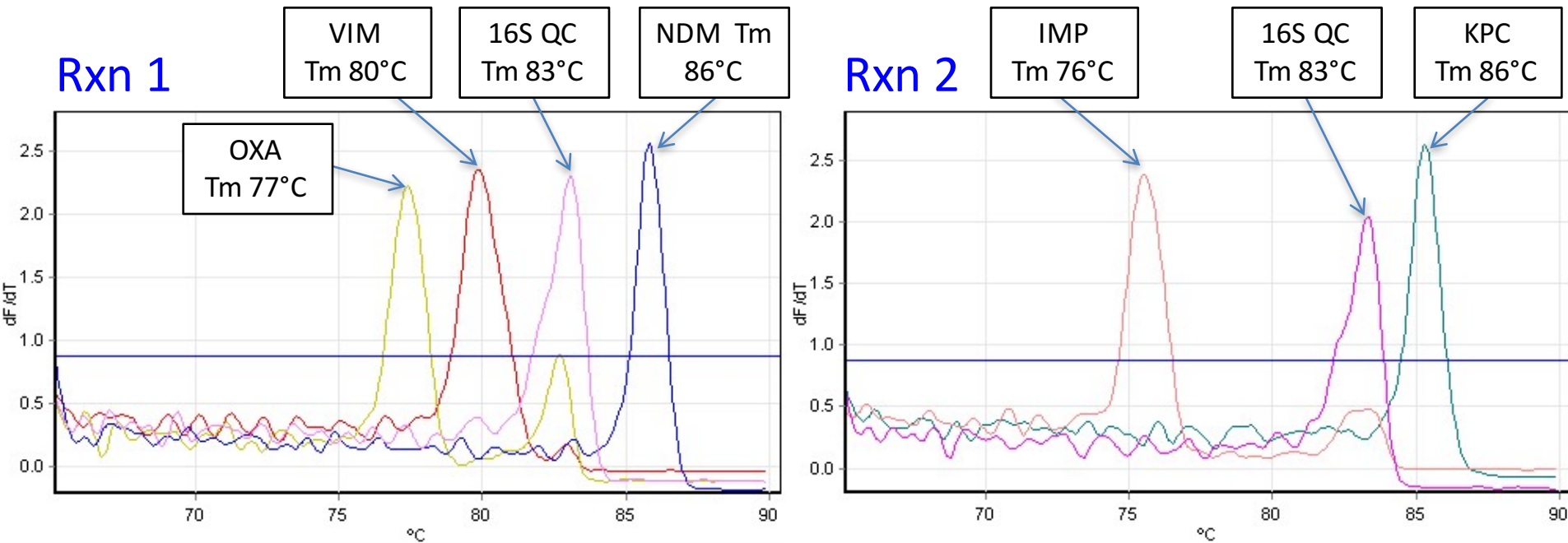
Specificity: 97.8%

Nucleic Acid Testing

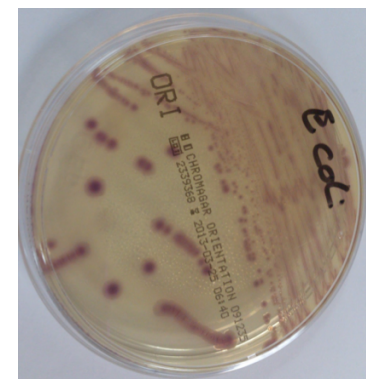


limited diversity of adjacent
Sequence suggests clonal spread

Stanford Multiplex CRE PCR



Indian man with pyelonephritis.
 Visiting daughter in Silicon Valley.
 Ureteral stent placed in India for kidney stones.
 Urine culture: *E. coli* >100,000/mL



Susceptibility

	MIC by Vitek 2		Disk Diffusion	MIC by Etest
Amikacin	>=64 ug/mL ...	RESISTANT		
Amoxicillin/Clavulanic Acid	>=32 ug/mL ...	RESISTANT		
Ampicillin	>=32 ug/mL ...	RESISTANT		
Cefazolin	>=64 ug/mL ...	RESISTANT ¹		
Cefepime	>=64 ug/mL ...	RESISTANT		
Cefoxitin	>=64 ug/mL ...	RESISTANT		
Ceftazidime	>=64 ug/mL ...	RESISTANT		
Ceftolozane/Tazobactam				>256 ug/mL ... R
Ceftriaxone	>=64 ug/mL ...	RESISTANT		
Ciprofloxacin	>=4 ug/mL (...)	RESISTANT		
Doxycycline				>256 ug/mL ... R
Ertapenem	>=8 ug/mL (...)	RESISTANT	RESISTANT	
FOSFOMYCIN			SUSCEPTIBLE	
Gentamicin	>=16 ug/mL ...	RESISTANT		
Imipenem			RESISTANT	
Levofloxacin	>=8 ug/mL (...)	RESISTANT		
Meropenem	8 ug/mL (MIC)	RESISTANT	RESISTANT	
Nitrofurantoin	64 ug/mL (MIC)	INTERMEDIATE		
Piperacillin/Tazobactam	>=128 ug/mL...	RESISTANT		
Polymixin B				1 ug/mL (MIC) No interpret
Tetracycline	>=16 ug/mL ...	RESISTANT		
Tobramycin	>=16 ug/mL ...	RESISTANT		
Trimethoprim/Sulfamethoxazole.	>=320 ug/mL...	RESISTANT		

Mechanism Guides Therapy for CRE

Enzyme	Class	Drug
KPC	A	Ceftazidime/Avibactam 100% Aztreonam/Avibactam 100%
IMP	B	Aztreonam/Avibactam 100%
VIM	B	Aztreonam/Avibactam 100%
NDM	B	Aztreonam/Avibactam 94%
OXA-48-like	D	Ceftazidime/Avibactam 93% Aztreonam/Avibactam 100%
None (Porin change +ESBL & or ampC)	A/C	Aztreonam/Avibactam: 97% Ceftazidime/Avibactam 100% Ceftolozane/Tazobactam ≈50%

 **NOW AVAILABLE**

Xpert® Carba-R:

On-demand molecular test detects and differentiates KPC, NDM, VIM, OXA-48 and IMP-1.

- *Rapid and accurate detection of carbapenemase-producing Gram-negative bacteria — with differentiation among the five major families of resistance genes*
- *Results in less than an hour to help guide implementation of epidemiological measures to control the spread of these microorganisms*
- *Fast, easy, and cost-effective, actionable information to help prevent widespread outbreaks*



  In Vitro Diagnostic Medical Device

FDA approved for culture

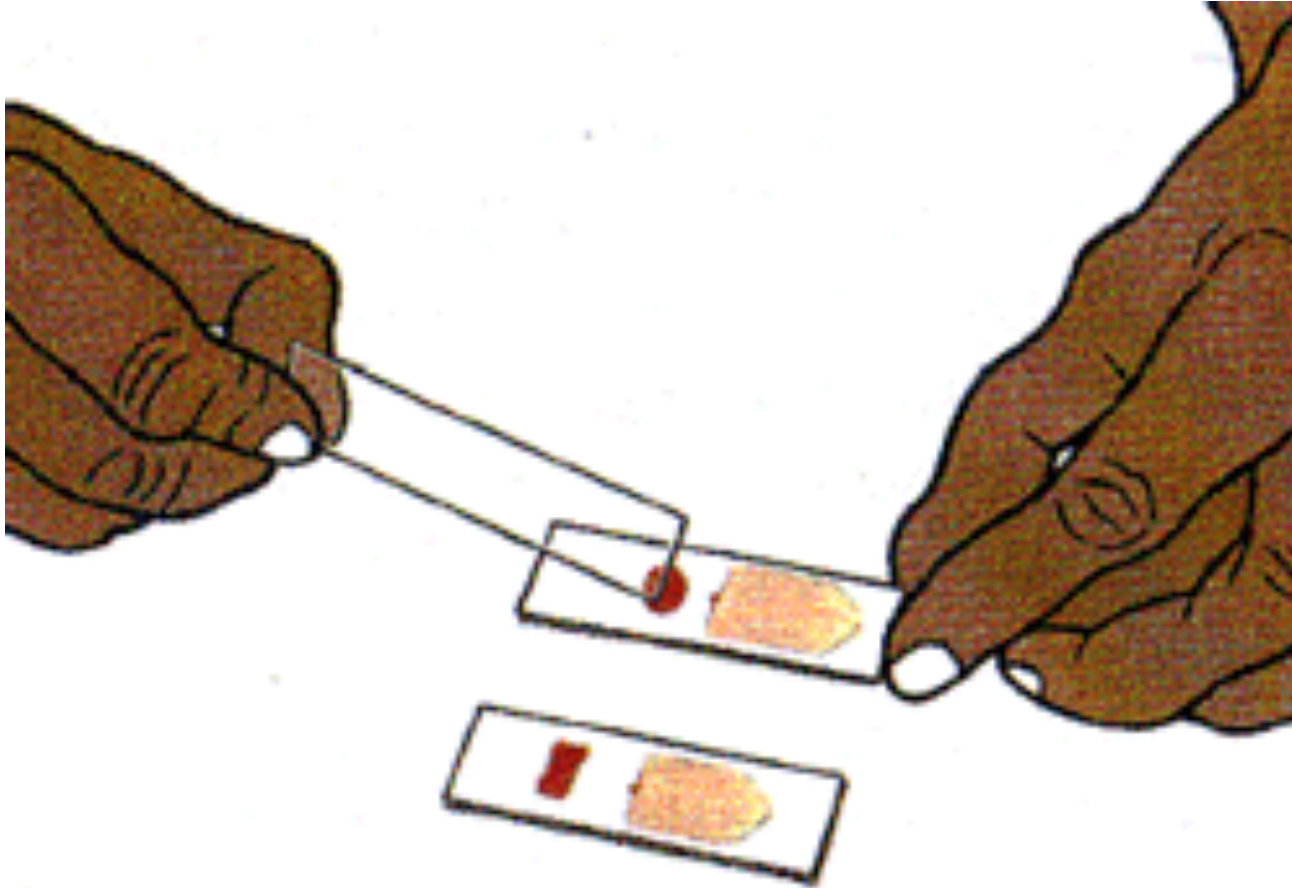
Disadvantage of PCR is that they can assess the presence of only already known resistance genes.

 **Cepheid**
A better way

Overview

- Accurate diagnostics in 21st century
- Diagnostics for emerging pathogens
 - Ebola and Zika
 - Carbapenem-Resistant Enterobacteriaceae (CRE)
 - Malaria

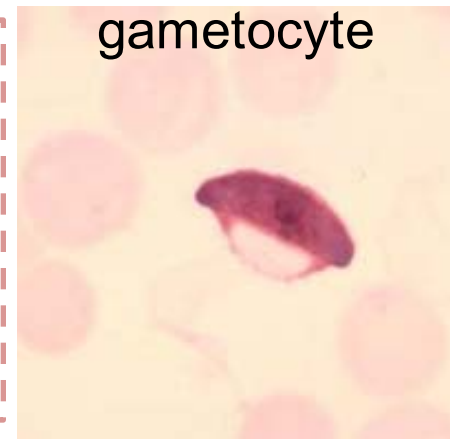
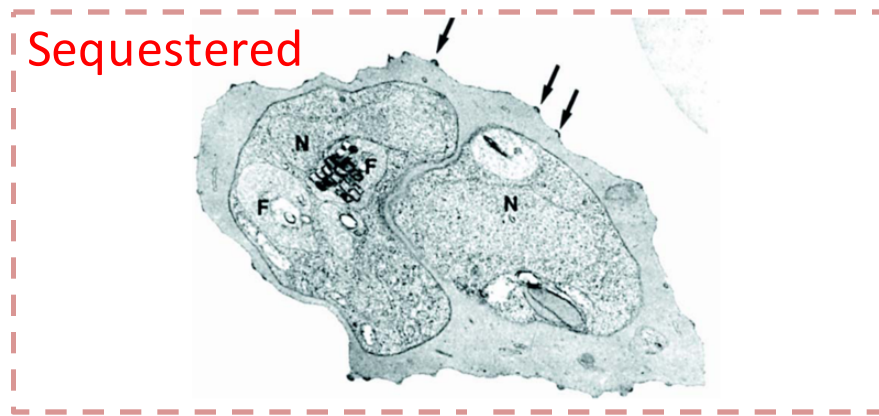
Diagnosis of Malaria by Light Microscopy



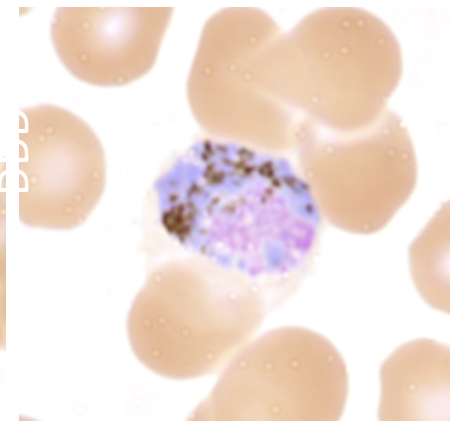
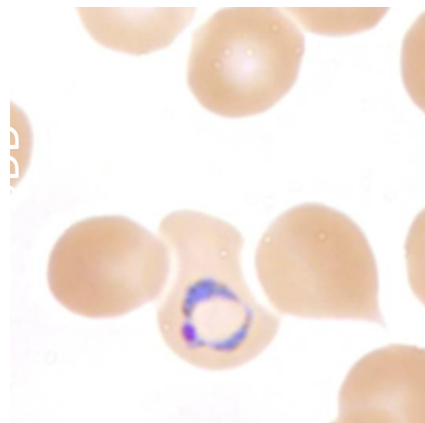
- Prepare slide immediately <1 hr after draw
- q8 h or q12 X 3 before malaria is r/o

Diagnosis of Malaria by Microscopy

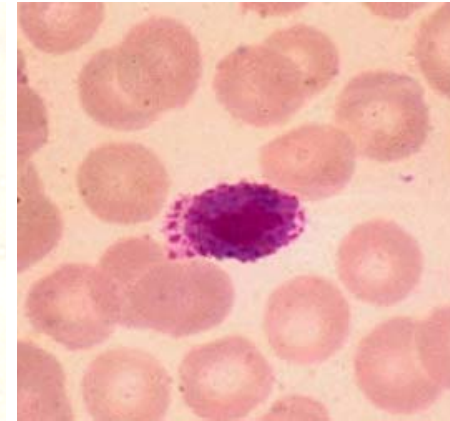
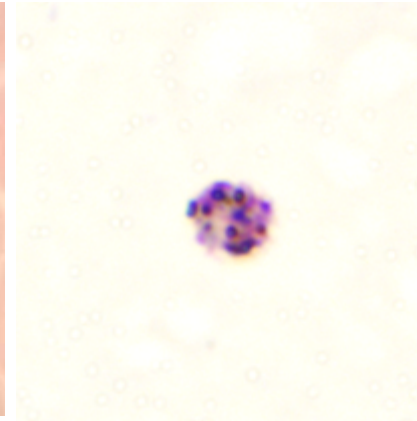
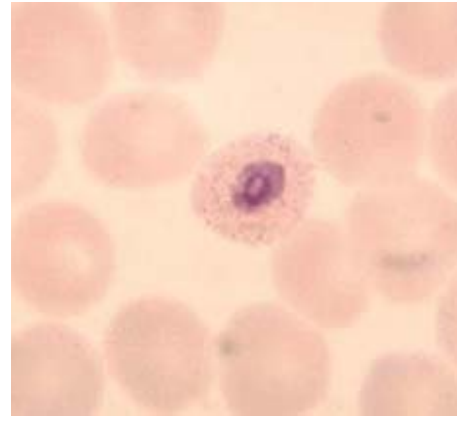
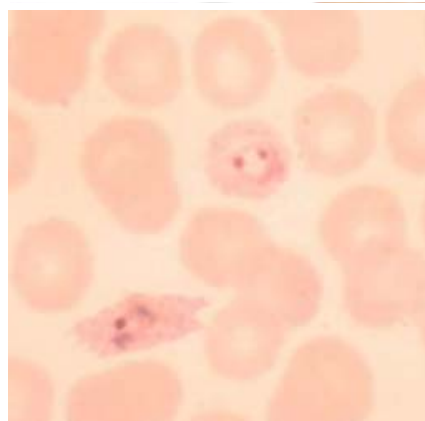
P.f



P.v

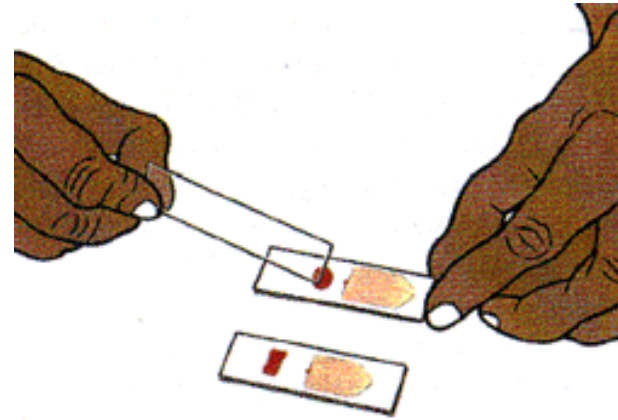


P.o

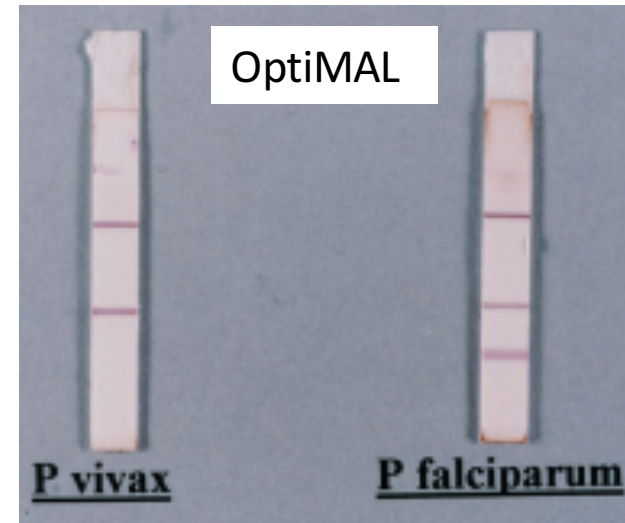
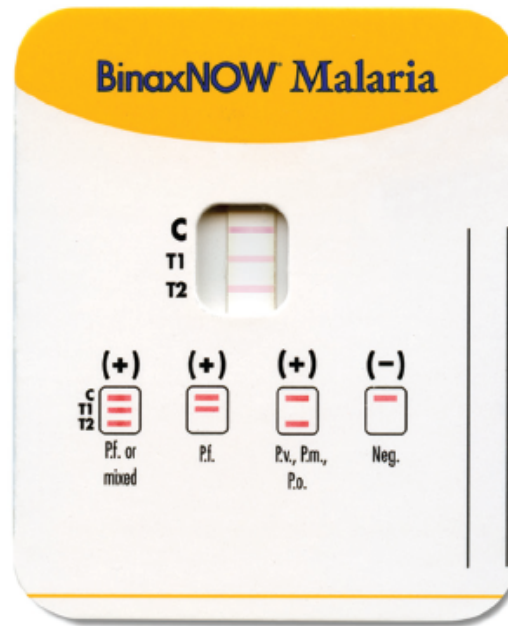
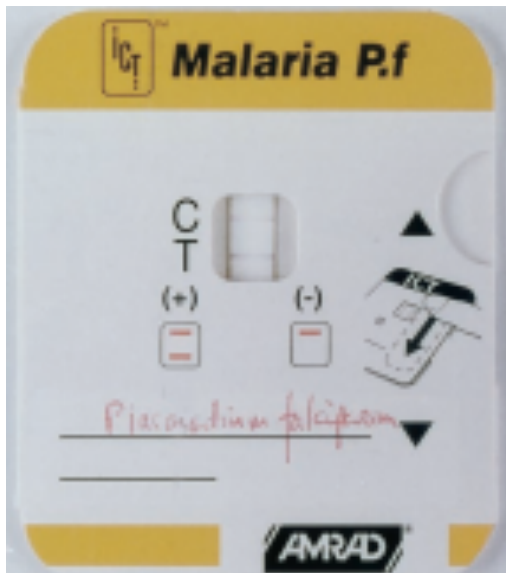


Light Microscopy (“Gold Standard”)

- Wreight-Giemsa stain cheap and durable
- Glass slides reusable
- Thin smear
 - Helpful for species identification
 - Allows % parasitemia
- Thic smear
 - More sensitive in diagnosing malaria (“Gold Standard”)
- Monitor therapy
 - Cure = 75% reduction in first 48h



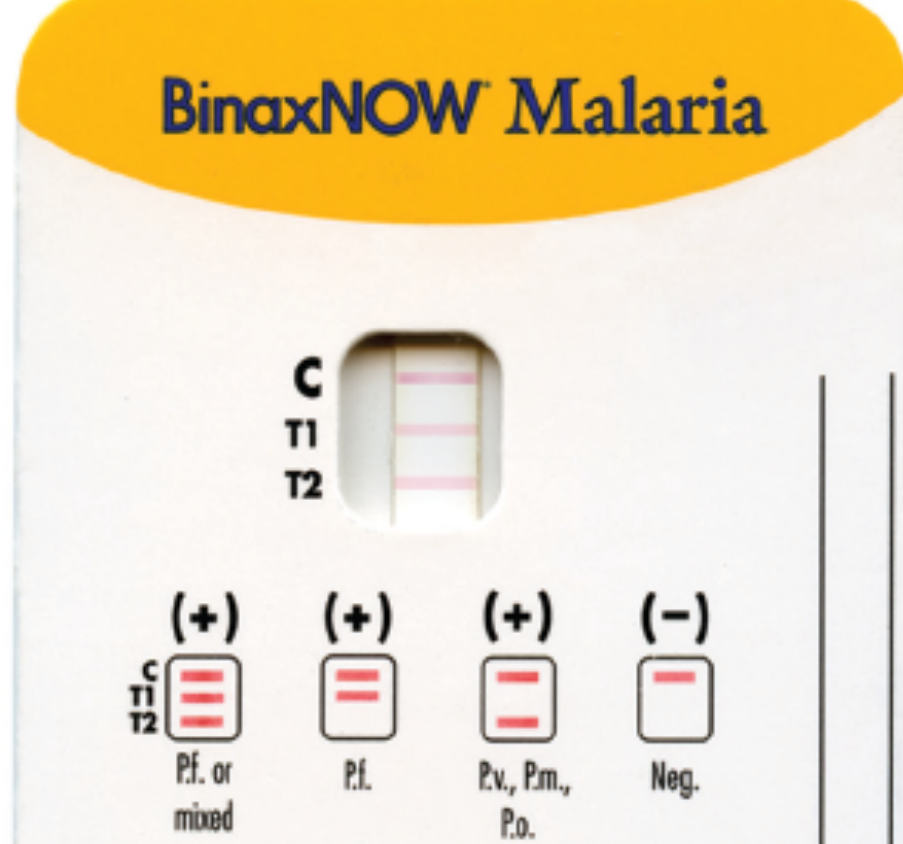
Nonmicroscopic Rapid Diagnostic Tests



Performance of Malaria Rapid Diagnostics in Endemic and Non-Endemic Areas

Reference	MRDD*†	Reference standard‡	Location	Blind study	Overall sensitivity (%)	Overall specificity (%)	<i>P. falciparum</i> sensitivity (%)	<i>P. falciparum</i> specificity (%)	<i>P. vivax</i> sensitivity (%)	<i>P. vivax</i> specificity (%)
<i>Endemic</i>										
Coleman et al. (2002b)	ICT Pf/Pv	GTTS	Thailand	Yes			35.4	99.7	2	99.9
Ferro et al. (2002)	OptiMAL	GTTS	Colombia	Unk§	99	94	90.6	98.6	96.5	97.6
Huong et al. (2002)	OptiMAL		Vietnam	Yes			50	100	73.7	100
	ICT Pf/Pv						83	100	20	100
Mason et al. (2002)	OptiMAL	GTTS	Myanmar	Yes	49		42	97	47¶	96.9¶
	ICT Pf/Pv				74		86	77	3¶	100¶
<i>Non-endemic</i>										
Gatti et al. (2002)	ICT Pf/Pv	GTTS	Italy	Unk	84	89	94	95	58¶	100¶
Iqbal et al. (2002)	OptiMAL	GTTS	Kuwait	Yes			87	99	79	97
	ICT Pf/Pv						81	99	58	98
Jelinek et al. (2001)	ICT Pf/Pv	GTTS	Europe	No			88	99	77	100
Playford & Walker (2002)	OptiMAL	GTTS and PCR	Australia	Yes	78	98	85	96	80	97
	ICT Pf/Pv				63	97	97	90	44	100
Rubio et al. (2001)	OptiMAL	GTTS and PCR	Europe	No	62	84				
	ICT Pf/Pv				68	97				

Limitations in sensitivity (100–500 parasites/ μ l), and robustness under field conditions.



BinaxNOW accuracy:

Plasmodium genus

Sensitivity is 92.9% (13/14)

Specificity is 99.8% (464/465)

Plasmodium species

P. falciparum 88.9% (8/9)

non-*falciparum* 100% (7/7)

Correlation with parasitemia

High parasitemia (1.0%, or 50,000/l): 100% (2/2)

Medium parasitemia (0.1 to 1.0%, or 5,000 to 50,000/l): 100% (12/12)

Low parasitemia (0.1%, or 5,000/l): 40% (2/5)

Limitations of Binax

Persistence of HRP-2 after parasite clearance

Humar et al. detected circulating HRP-2 antigen in 68% of treated patients on day 7, and in **27% it was still present on day 28** (Am J Trop Med Hyg 1997 **56:44**)

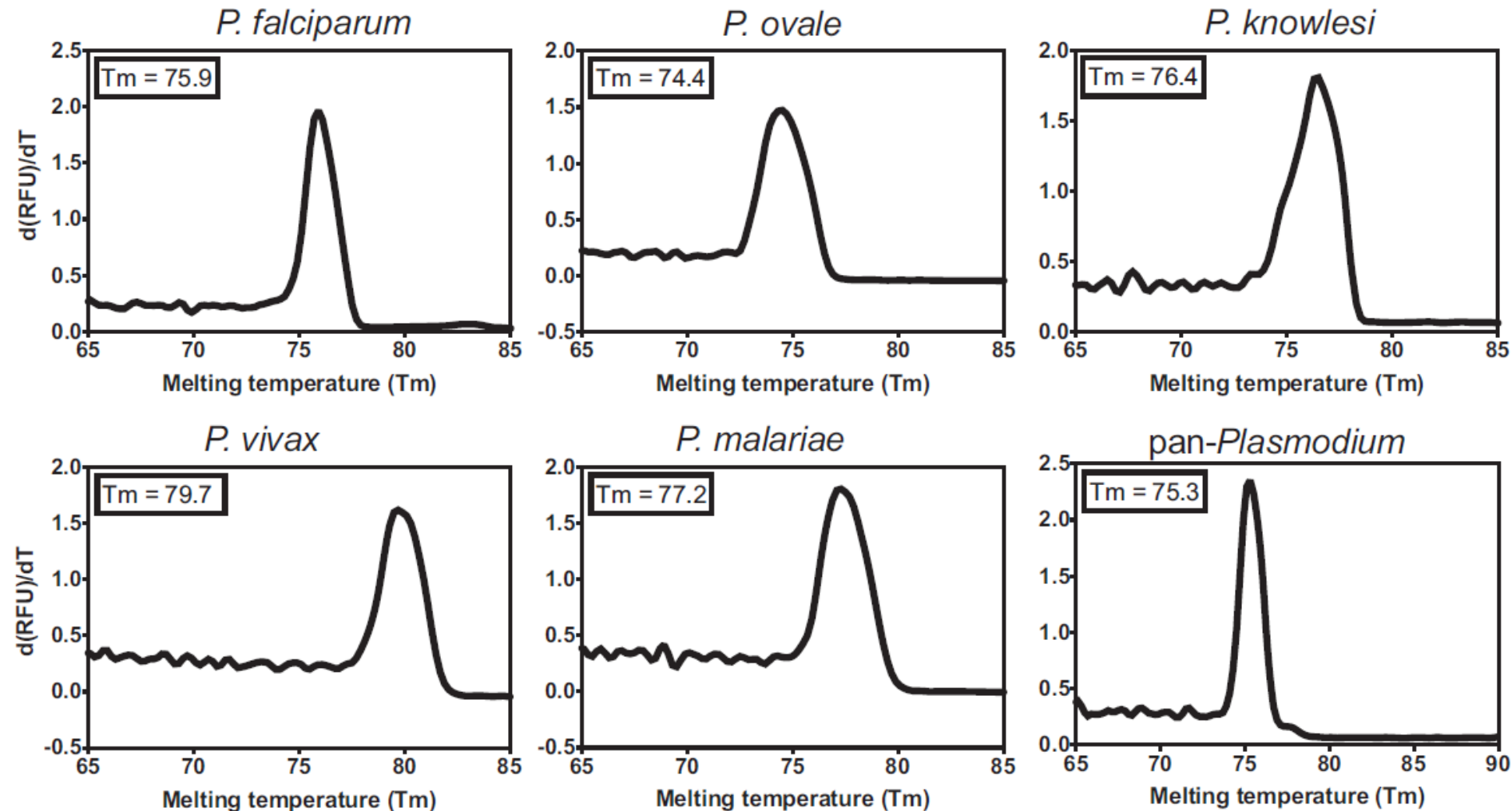
Gene deletion for the production of HRP-2

70% in Peru (per Joe Vinetz at UCSD)

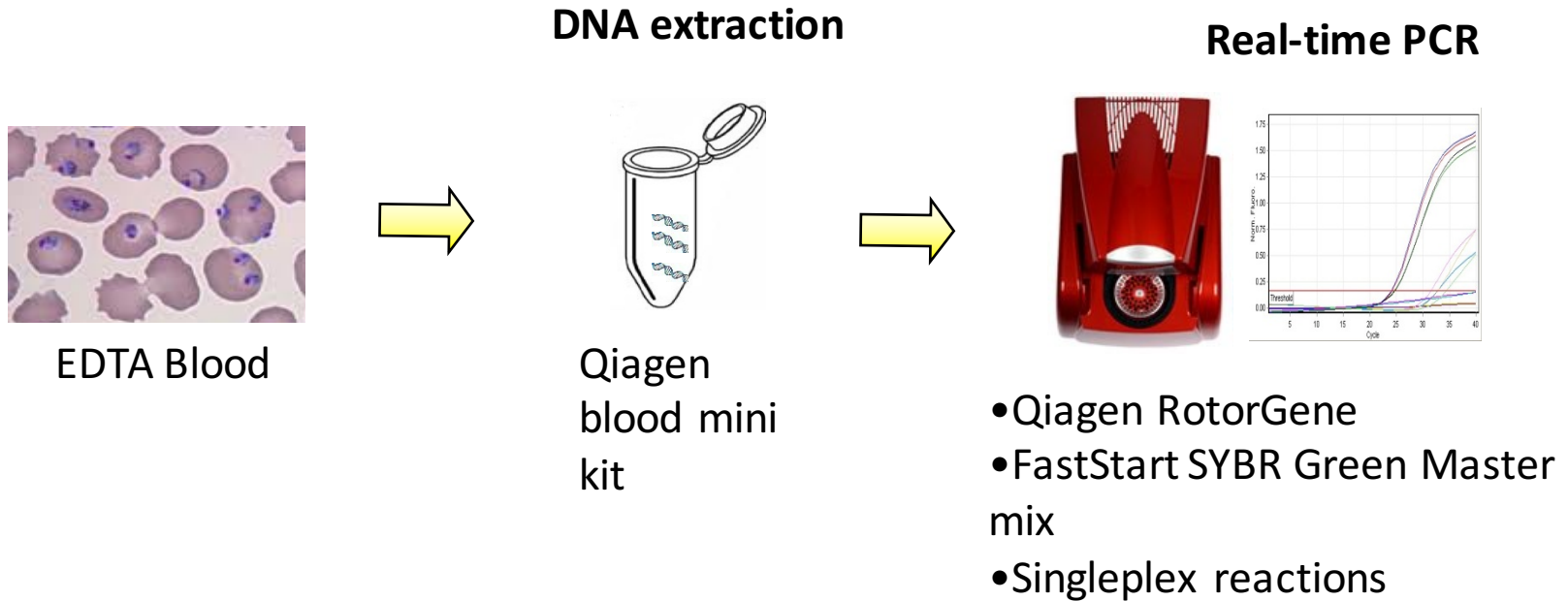
False-positive results

Cross-reaction with serum rheumatoid factor

Malaria Detection and Species Identification by Real-Time PCR



Malaria Detection and Species Identification by Real-Time PCR



Diagnostics No Longer a Limiting Factor

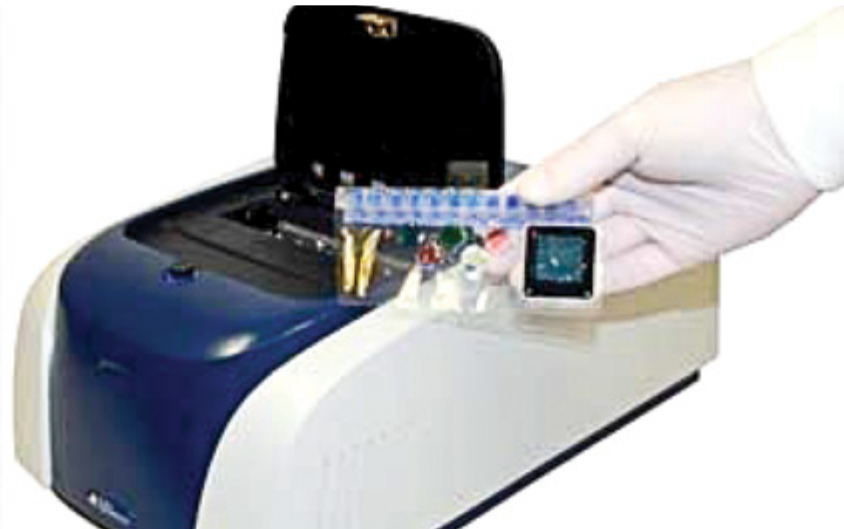
GeneXpert Omni, Cepheid



IMMY



Biofire, bioMérieux

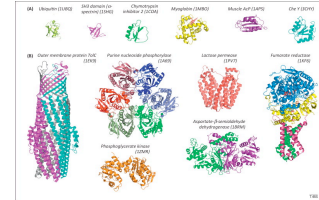
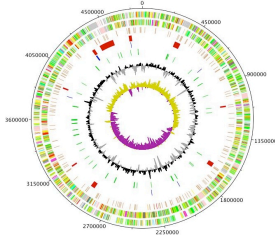


Cobas Liat System, Roche



Summary I

Genomics/ Proteomics Databases

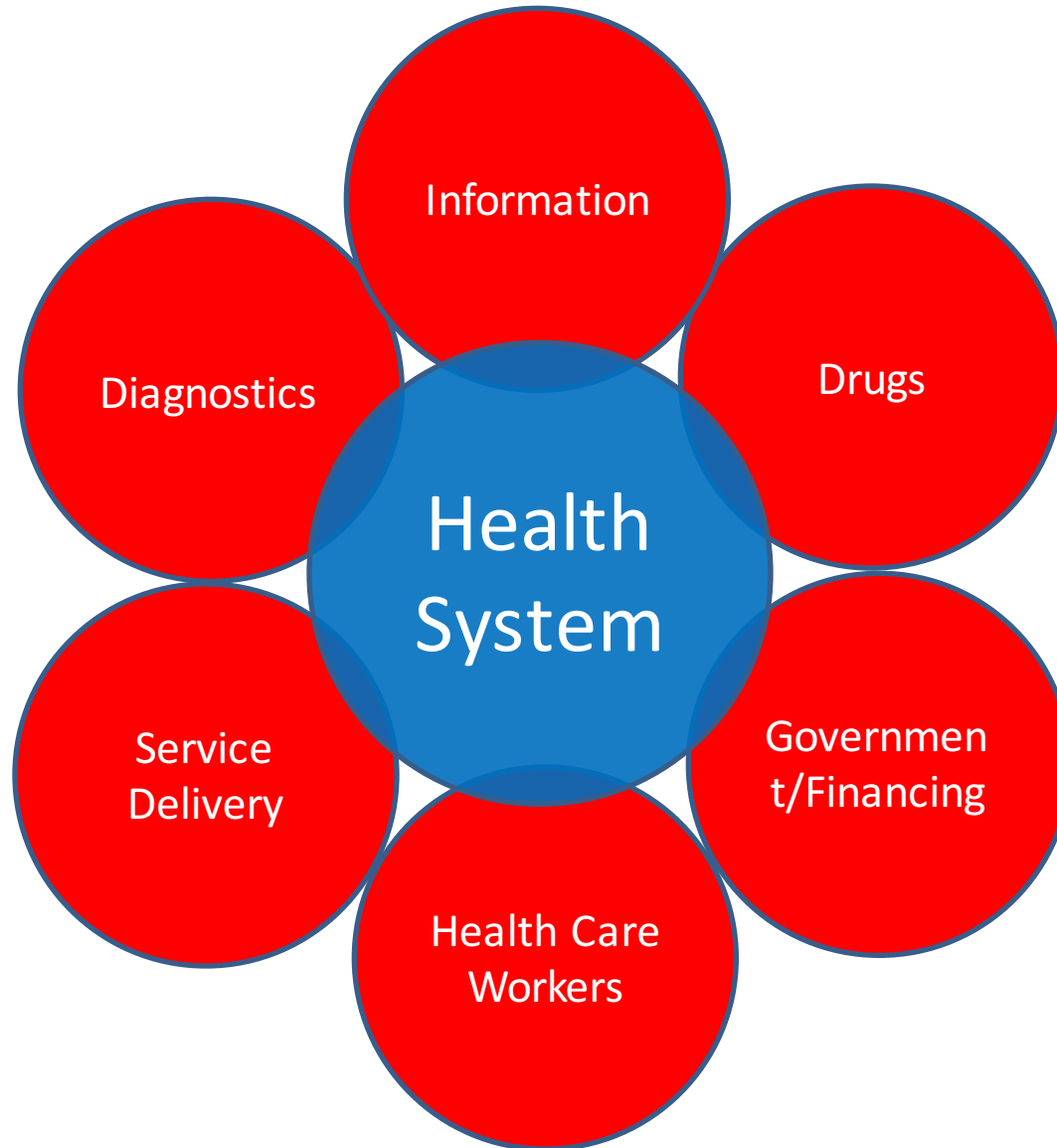


Molecular
Diagnostic
Platforms



Bioinformatics
Tools

Summary II



Acknowledgements

Stanford University

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Jana Broadhurst

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Stanford Pathology

Merck Inc

CLSI Interpretive Guidelines

Table 2. Breakpoint values (MIC, mg/L) for carbapenems according to guidelines in Europe (EUCAST) and the United States (CLSI), September 2010*

Carbapenem	EUCAST		CLSI	
	S	R	S	R
Ertapenem	<u>≤</u> 0.5	>1	<u>≤</u> 0.25	<u>≥</u> 1
Imipenem	<u>≤</u> 2	>8	<u>≤</u> 1	<u>≥</u> 4
Meropenem	<u>≤</u> 2	>8	<u>≤</u> 1	<u>≥</u> 4

*EUCAST, European Committee on Antimicrobial Susceptibility Testing (www.eucast.org/clinical_breakpoints); CLSI, Clinical and Laboratory Standards Institute; S, sensitive; R, resistant.

Urgent Need for Measures to Contain Carbapenemase-Producing Enterics

- Complete and continuous epidemiological data
 - Global data: systematic reports from Africa, the Balkans, the Middle East, and vast areas in Asia
 - Multi-institution data from developed countries
- Implementation of effective yet affordable and sustainable measures against CPE spread
- Identifying the most effective drug combinations
 - Dosing regimen of colistin, carbapenems
- Resources from international public health organizations should be mobilized and allocated appropriately

TABLE 4. Substrate and inhibition profiles of the carbapenemases

Molecular class	Functional group	Enzyme	Hydrolysis profile ^a					Inhibition profile ^b		Reference(s)
			Penicillins	Early cephalosporins	Extended-spectrum cephalosporins	Aztreonam	Carbapenems	EDTA	Clavulanic acid	
A	2f	NMC	+	+	+	+	+	—	+	124
		IMI	+	+	+	+	+	—	+	183
		SME	+	+	±	+	+	—	+	179
		KPC	+	+	+	+	+	—	+	4
		GES	+	+	+	—	±	—	+	174, 219
B1	3	IMP	+	+	+	—	+	+	—	224
		VIM	+	+	+	—	+	+	—	224
		GIM	+	+	+	—	+	+	—	224
		SPM	+	+	+	—	+	+	—	224
D	2d	OXA	+	+	±	—	±	—	±	225

Biochemical Identification of Enzymes

➤ KPC

- Boronic acid-based inhibition testing
- Performed with imipenem or meropenem

➤ NDM, VIM, IMP

- Etest MBL strip (bioMérieux)
- EDTA inhibits MBL activity

➤ OXA-48-like

- No inhibition test is available

Mechanism Guides Therapy for CRE

Enzyme	Class	Drug
KPC	A	Ceftazidime/Avibactam 100% Aztreonam/Avibactam 100%
IMP	B	Ceftazidime/Avibactam 9% Aztreonam/Avibactam 100%
VIM	B	Ceftazidime/Avibactam 0% Aztreonam/Avibactam 100%
NDM	B	Ceftazidime/Avibactam 0% Aztreonam/Avibactam 94%
OXA-48-like	D	Ceftazidime/Avibactam 93% Aztreonam/Avibactam 100%
None (Porin change +ESBL & or ampC)	A/C	Aztreonam/Avibactam: 97% Ceftazidime/Avibactam 100% Ceftolozane/Tazobactam ≈50%

Mechanism Guides Therapy for CRE

Enzyme	Class	Drug
KPC	A	Ceftazidime/Avibactam Aztreonam/Avibactam
IMP	B	Aztreonam/Avibactam
VIM	B	
NDM	B	
OXA-48-like	D	Ceftazidime/Avibactam Aztreonam/Avibactam
Porin +ESBL/ampC	A/C	Ceftazidime/Avibactam Aztreonam/Avibactam

Among 108 KPC, 32 NDM, 11 IMP, 14 OXA-48-like, 5 IMI, 4 VIM, and 3 SME producers:
aztreonam-avibactam active against all except two NDM with ↑MICs of 8/4 & 16/4 mg/L;
ceftazidime-avibactam active against all KPC, IMI, SME, and most OXA-48-like (93%) but not class B

Among 29 enterics: 18 ESBL (10 porin loss), 11 AmpC (3 porin loss), 1 ESBL+AmpC:
aztreonam-avibactam active against all except one (8/4 mg/L); ceftazidime-avibactam active against all

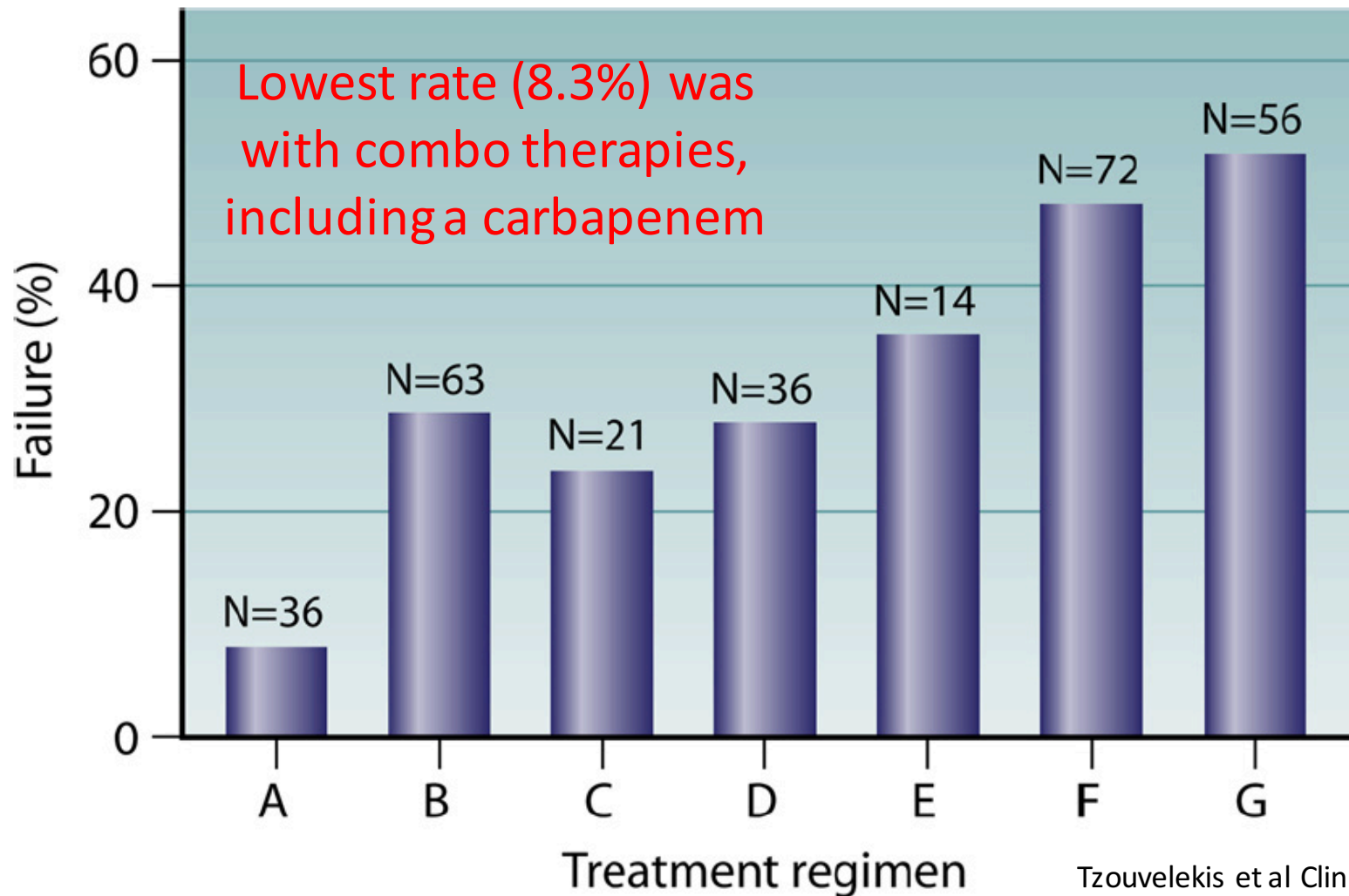
Older antibiotics: colistin 88%; tigecycline 79%; and fosfomycin 78%

Vasoo et al AAC 2016

TABLE 3 Comparative susceptibilities of carbapenemase-producing Gram-negative bacilli to various antimicrobials

Antimicrobial ^a	% susceptibility with resistance mechanism shown (total no. of isolates)						
	KPC (108)	NDM (32)	OXA-48 group (14)	IMP (11)	IMI (5)	VIM (4)	SME (3)
β-Lactams and β-lactam–β-lactamase inhibitor combinations							
ATM	2	9	14	18	100	100	0
AZA	100	94	100	100	100	100	100
CAZ	2	0	14	0	80	0	67
CZA	100	0	93	9	100	0	100
CRO ^b	1	0	14	0	100		67
FEP ^c	2	0	14	0	80	0	33
ETP ^b	2	0	7	0	0		0
MEM	5	0	43	36	20	0	0
TZP	1	0	0	27	100	0	67
MEC ^b	0	16	43	18	100		33
Aminoglycosides							
GEN	63	16	36	0	80	0	100
TOB	9	9	36	9	80	0	100
AMK	51	41	43	82	100	0	100
Fluoroquinolones							
LVX	11	13	14	36	100	0	100
CIP	10	0	7	18	80	0	67
Other antimicrobial agents							
SXT ^b	21	9	14	45	80		100
NIT ^b	14	25	7	27	20		
FOF ^b	78	78	71	73	100		100
CST ^d	87	91	100	91	40	75	
TGC ^{b,d}	81	69	93	55	100		100

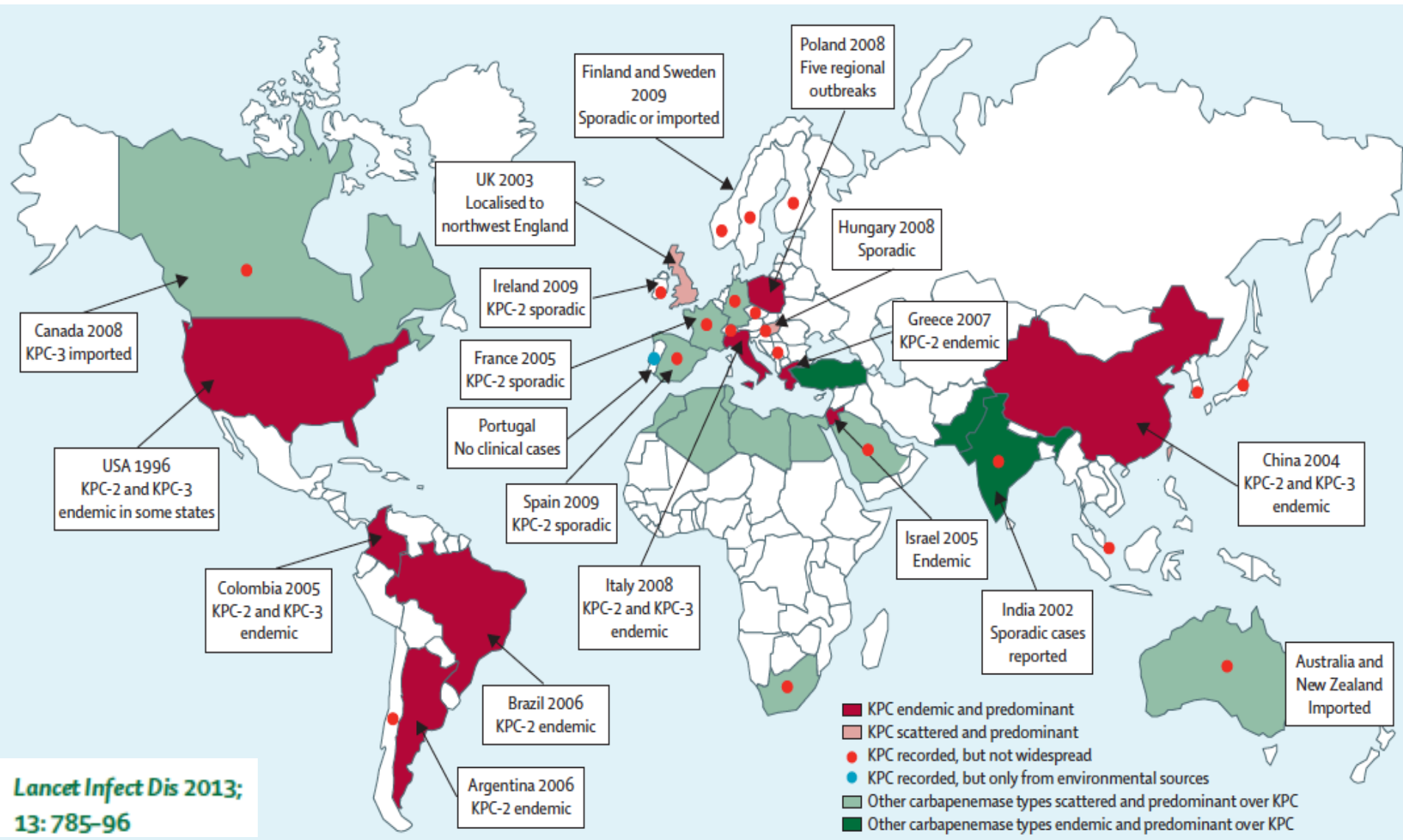
Clinical Outcomes of Carbapenemase-Producing *Klebsiella pneumoniae*



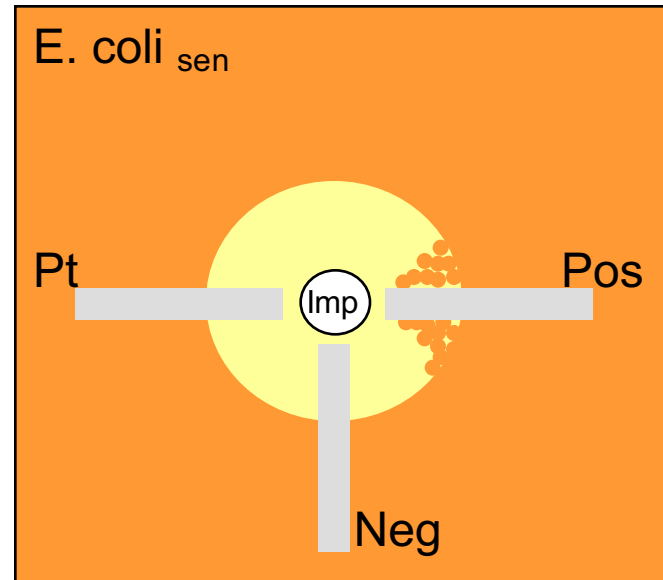
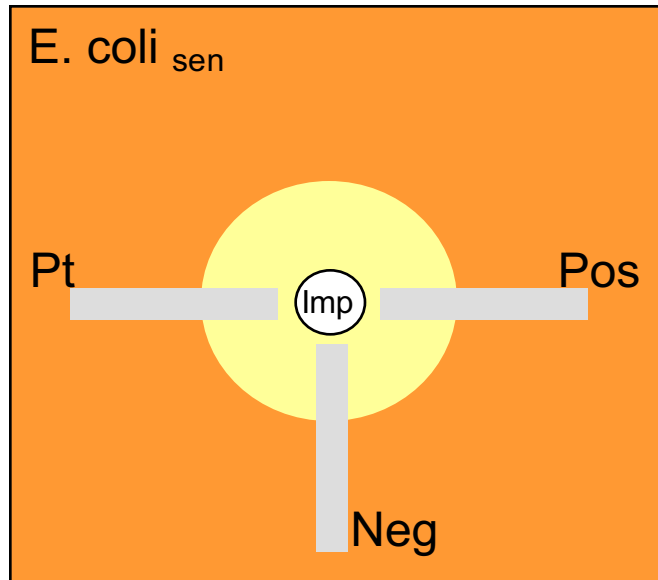
Tzouveleakis et al Clin Micro Rev 2012

A, combination tx with 2 active drugs, one was a carbapenem; B, combination tx with 2 active drugs, not including a carbapenem; C, monotherapy with an aminoglycoside; D, monotherapy with a carbapenem; E, monotherapy with tigecycline; F, monotherapy with colistin; G, inappropriate tx.

Geographic Distribution of *Klebsiella pneumoniae* Carbapenemase (KPC) producers



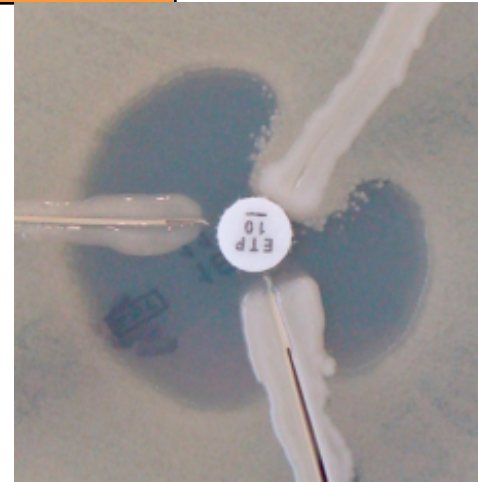
Phenotypic Detection of Carbapenemases: Modified Hodge Test



Sensitivity: 98% (weak detection of NDM producers)
Specificity: 80% (high level AmpC producers)

Vasoo et al JCM 2013

Nordmann et al Emerg Infect Dis 2011



Will Ebola change the game? Ten essential reforms before the next pandemic. The report of the Harvard-LSHTM Independent Panel on the Global Response to Ebola



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Panel 1: Summary of system weaknesses exposed across four phases of the Ebola outbreak

Pre-outbreak and Phase 1: December, 2013–March, 2014

- Inadequate national investment and donor support for building national health systems capable of detecting and responding to disease outbreaks
- Inadequate arrangements to monitor country commitments to do so

Phase 2: April, 2014–July, 2014

- Little incentive for countries to report outbreaks early
- Insufficient overall technical capacity among national and international teams
- WHO slow to mobilise global attention or assistance

Phase 3: August, 2014–October, 2014

- Government and private sector disregard for WHO recommendations regarding travel and trade restrictions
- Slow global operational response
- Unclear responsibility for international coordination
- Weak channels for lessons from previous Ebola outbreaks
- Little access to therapies in limited supply, or medical evacuation for national as opposed to international health workers
- Poor understanding of the importance of community engagement

Phase 4: October, 2014–September, 2015

- Weak coordination of global operational response
- Inadequate transparency on resource flows
- Weak accountability for use of funds at all levels